









FISH SCALES HYDROXYAPATITE/COLLAGEN COMPOSITES INCORPORATED WITH SILVER NANOPARTICLES FOR BONE FILLER **APPLICATIONS**

MUSTAFA MUDHAFAR JAHIL











THESIS SUBMITTED IN FULFILLMENT OF THE REQUIREMENT FOR THE DEGREE OF DOCTOR OF PHILOSOPHY

FACULTY OF SCIENCE AND MATHEMATICS SULTAN IDRIS EDUCATION UNIVERSITY

2021

















UPSI/IPS-3/BO 32 Pind: 00 m/s: 1



ii



INSTITUTE OF GRADUATE STUDIES DECLARATION OF ORIGINAL WORK

Please tick ($$):	
Project Paper	
Masters by Research	
Master by Mixed Mode	
PhD	V

This declaration is made on the 12 / 1 /2021

Student's Declaration:

MUSTAFA MUDHAFAR JAHIL (P20162002431) FACULTY OF SCIENCE AND _hereby declare that the work entitled FI\$H SCALES COMPOSITES INCORPORATED WITH HYDROXYAPATITE/COLLAGEN ____ NANOPARTICLES FOR BONE FILLER APPLICATIONS is my original work. I have not copied from any other students' work or from any other sources except where due reference or acknowledgement is made explicitly in the text, nor has any part been written for me by another person.

Signature of the student

ii. Supervisor's Declaration:

I, PROF. DR. ISMAIL ZAINOL hereby certifies that the work entitled FISH SCALES HYDROXYAPATITE / COLLAGEN COMPOSITES INCORPORATED WITH SILVER NANOPARTICLES FOR BONE FILLER APPLICATIONS was prepared by the above named student, and was submitted to the Institute of Graduate Studies as a fulfillment for the conferment of PH.D. IN CHEMISTRY, and the aforementioned work, to the best of my knowledge, is the said student's work.













iii



INSTITUT PENGAJIAN SISWAZAH / INSTITUTE OF GRADUATE STUDIES

BORANG PENGESAHAN PENYERAHAN"[TESIS/DISERTASI/LAPORAN KERTAS PROJEK]" DECLARATION OF "[THESIS/DISSERTATION/PROJECT PAPER FORM]"

Tajuk / Title: FISH SCALES HYDROXYAPATITE/COLLAGEN COMPOSITES INCORPORATED WITH SILVER NANOPARTICLES FOR BONE FILLER APPLICATIONS

No. Matrik / Matric's No.: P20162002431

Saya / 1: **MUSTAFA MUDHAFAR JAHIL**

mengaku membenarkan [Tesis/Disertasi/Laporan-Kertas-Projek] (Doktor-Falsafah/Sarjana)* ini disimpan di Universiti Pendidikan Sultan Idris (Perpustakaan Tuanku Bainun) dengan syaratsyarat kegunaan seperti berikut:-

acknowledged that my [Thesis/Dissertation/Project-Paper] is kept at Universiti Pendidikan Sultan Idris (Tuanku Bainun Library) and reserves the right as follows:-

1. Tesis/Disertasi/Laporan Kertas Projek ini adalah hak milik UPSI. The thesis is the property of Universiti Pendidikan Sultan Idris

2. Perpustakaan Tuanku Bainun dibenarkan membuat salinan untuk tujuan rujukan sahaja. Tuanku Bainun Library has the right to make copies for the purpose of research only.

3. Perpustakaan dibenarkan membuat salinan Tesis/Disertasi ini sebagai bahan pertukaran antara Institusi Pengajian Tinggi.

The Library has the right to make copies of the thesis for academic exchange.

4. Perpustakaan tidak dibenarkan membuat penjualan salinan Tesis/Disertasi ini bagi kategori TIDAK TERHAD.

The Library is not allowed to make any profit for 'Open Access' Thesis/Dissertation.

5. Sila tandakan (√) bagi pilihan kategori di bawah / Please tick (√) for category below:-

SULITICONFIDENTIAL	. Mengandungi maklumat yang berdarjah keselamatan atau kepentingan Malaysia seperti yang termaktub dalam Akta Rahsia Rasmi 1972. I Contains confidential
	information under the Official Secret Act 197
TERHAD/RESTRICTE	D Mengandungi maklumat terhad yang telah ditentukan oleh organisasi/badan di mana penyelidikan ini dijalankan. / Contains restircted information as specified by
	the organization where research was done.

TIDAK TERHAD I QPEN ACCESS

Tarikh: 2611/2021

NU Tandatangan-Pelajar/ Signature) (Tandatangan Penyellabatan Inter Supervisor) Fakulti Sains dan Matematik

&(Nama & Copiessili Asadidikan Sultan Hari

Catatan: Jika Tesis/Disertasi ini SULIT @ TERHAD, sila lampirkan surat daripada pihak berkuasa/organisasi berkenaan dengan menyatakan sekali sebab dan tempoh laporan ini perlu dikelaskan sebagai SULIT dan TERHAD.

Notes: If the thesis is CONFIDENTAL or RESTRICTED, please attach with the letter from the organization with period and reasons for confidentiality or restriction.

















iv

ACKNOWLEDGEMENTS

I thank God, my helper, my shield, my defence, my stronghold and the maker of all things because, without Him, this research could not have been concluded. Thank you, my God.

I would like to express my deepest gratitude to my main supervisor, Professor Dr Ismail Zainol, and Co- supervisors, Dr. Che Nor Aiza Binti Jaafar from Universiti Putra Malaysia (UPM) for their scholarly support, academic guidance, encouragement, understanding, invaluable assistance, constructive criticism and insight at all times during my study. Their guidance has provided me with useful critical comments, suggestions and strong support for doing the research which has shaped my ideas and thesis structure.

I would like to thanks the academic and non-academic staff of the faculty of science and mathematics in UPSI for providing generous assistance, valuable and useful suggestions during my time in this university. I also thank all members of faculty in the Department of Chemistry for their support throughout the duration of this work.

Last but not least, I owe a great debt to my family; my father, my mother, my brothers and sisters for their support throughout the duration of this study and I would like to express my appreciation to many friends and colleagues for their interest, enjoyment, **O** ptbupsi encouragement and valuable support throughout my study.



















ABSTRACT

The aim of this study was to improve antimicrobial properties of fish scales hydroxyapatite/fish collagen (FsHA/FsCol) composites through infiltration of silver nanoparticles into the matrix for antimicrobial bone filler substitute. In this sudy both hydroxyapatite and collagen were extracted from fish scales using simple hydrothermal extraction method. Silver nanoparticles (AgNPs) were synthesized from silver nitrate using microwave-assisted method in which neems leaves extract was used as reducing agent while fish collagen as stabilizer. FsHA/FsCol/AgNPs composites were prepared in a form of powders and beads using two different methods. Chemical and physical properties of AgNPs and FsHA/FsCol/AgNPs composites were characterized using field emmision scanning electron microscope with energy dispersive x-ray (FESEM-EDX), transmission electron microscopy (TEM), Fourier transform infrared spectroscopy (FTIR) and x-ray diffraction technique (XRD). Biological properties of prepared composites were investigated through antibacterial, cytotoxicity and cell attachment tests. The results shown that four different sizes for AgNPs ranging from 28 nm to 100 nm were obtained by using different concentration of collagen. Their antimicrobial activities against Escherichia coli and Staphylococcus aureus showed that smaller particles sizes of AgNPs have shown better inhibition of bacteria growth. Analysis using XRD and EDX proved that the pattern of spectra corresponded to silver nanoparticles present in the composites. FESEM-EDX analysis indicated that the silver nanoparticles distributed wellin the HA composite matrix. Composites with 80 wt.% of HA and 20 wt.% of collagen demonstrated high stability during degradation test. The addition of 20 wt.% starch in the composite during preparation improved the porosity of composites after sintering, thus improved the infiltration of AgNPs-collagen solution and antibacterial properties. In conclusion, composites of FsHA/FsCol/AgNPs powder and beads have been proved to be biocompatible and good cell attachment. The implication of this study is both powder and beads of FsHA/FsCol/AgNPs composties can be used as antimicrobial bone filler substitute.





















KOMPOSIT HIDROKSIAPATIT/KOLAGEN SISI IKAN DIGABUNG DENGAN NANOPARTICLES SILVER UNTUK APLIKASI PENGISI TULANG **ANTIMIKROBIAL**

ABSTRAK

Tujuan kajian ini adalah untuk meningkatkan sifat antimikrobial komposit hidroksiapatit/kolagen (FsHA/FsCol) sisik ikan melalui penyusupan nanopartikel perak ke dalam matriks sebagai pengganti pengisi tulang antimikrobial. Dalam kajian ini kedua hidroksiapatit dan kolagen diekstrak dari sisik ikan menggunakan kaedah pengekstrakan hidrotermal. Nanopartikel perak (AgNPs) disintesis dari perak nitrat menggunakan kaedah berbantukan gelombang mikro di mana ekstrak daun mambu digunakan sebagai penurunan manakala kolagen ikan sebagai agen penstabil. FsHA/FsCol/AgNPs disediakan dalam bentuk serbuk dan manik menggunakan dua kaedah yang berbeza. Sifat kimia dan fizikal AgNPs dan komposit FsHA/FsCol/AgNP dicirikan menggunakan mikroskop pengimbas elektron pelepasan medan dengan penyebaran tenaga sinar-x (FESEM-EDX), spektroskopi inframerah transformasi Fourier ptbupsi (FTIR), mikroskop transmisi elektron (TEM) dan teknik pembelauan sinar-x (XRD). Sifat biologi komposit yang dihasilkan dikaji melalui ujian antibakteria, sitotoksisiti dan lekatan sel. Hasil kajian menunjukkan empat AgNPs yang berbeza saiz antara 28 nm hingga 100 nm dihasilkan dengan menggunakan kepekatan kolagen yang berbeza. Kegiatan antimikrobial komposit terhadap Escherichia coli dan Staphylococcus aureus telah menunjukkan bahawa ukuran partikel AgNP yang lebih kecil menunjukkan penghalangan pertumbuhan bakteria yang lebih baik. Analisis menggunakan XRD dan EDX telah membuktikan corak spektra sesuai dengan kehadiran nanopartikel perak yang terdapat di dalam komposit. Analisis FESEM-EDX telah menunjukkan bahawa nanopartikel perak tersebar dengan baik dalam matriks komposit HA. Komposit dengan 80% berat HA dan 20% berat kolagen telah menunjukkan kestabilan yang tinggi semasa ujian degradasi. Penambahan 20% berat tepung ubi dalam komposit semasa penyediaan telah meningkatkan keliangan komposit selepas pensinteran, sehingga meningkatkan penyusupan larutan AgNPs-kolagen dan seterusnya meningkatkan sifat antibakteria. Kesimpulannya, komposit serbuk dan manik FsHA/FsCol/AgNPs terbukti bioserasi dan menunjukkan lekatan sel yang baik. Implikasi dari kajian ini adalah serbuk dan manik dari komposit FsHA/FsCol/AgNPs boleh digunakan sebagai pengganti pengisi tulang antimikrobial.















vii

CONTENTS

		Page		
DECLARATIO	N OF ORIGINAL WORK	ii		
DECLARATIO	N OF THESIS	iii		
ACKNOWLED	GEMENTS	iv		
ABSTRACT		v		
ABSTRAK		vi		
TABLE OF CO	NTENTS	vii		
LIST OF FIGUE	RES	xv		
LIST OF TABLES xvii				
LIST OF ABBREVIATIONS Perpustakaan Tuanku Bainun Kampus Sultan Abdul Jalil Shah PustakaTBainun XXII Ptbups				
CHAPTER 1	INTRODUCTION	1		
1.1	Background of study	1		
1.2	Problem Statement	8		
1.3	Research Objectives	10		
1.4	Significance of study	11		
CHAPTER 2	LITERATURE REVIEW	12		
2.1	Bone	12		
	2.1.1 Bone defect	13		
	2.1.2 Bone healing process	14		
	2.1.3 Rone graft materials	14		











				V111
2.2	Hydro	oxyapatit	e	23
		2.2.1	Synthetic of HA	24
		2.2.2	Natural Hydroxyapatite	26
	2.3	Collage	en in bone repair	29
		2.3.1	Pure Collagen	29
		2.3.2	Fish scals Collagen	31
	2.4	Collage	n-hydroxyapatite composites	34
	2.5	Beads o	of Collagen/ Hydroxyapatite	42
	2.6	Silver	and Silver nanoparticles	42
	2.7	Applica	tions of Silver nanoparticles	43
	2.8	Antibac	eterial properties of AgNPS	45
) 05-450683		2.8.1	Effect of silver nanoparticle shapes on the inhibition zone of bacterial	48 ptbups
		2.8.2	Effect of silver nanoparticle sizes on the inhibition zone of bacterial	49
		2.8.3	Effect of silver nanoparticle concentrations on inhibition zone of bacterial	51
	2.9	Escher	ichia coli and Staphylococcus aureus	52
	2.10	Mechar	nism action of silver nanoparticles on bacteria	53
	2.16	Cytotox	cicity of silver nanoparticles	55
	2.12	Method	Is to synthesis nanoparticles	59
		2.12.1	Physical methods of silver nanoparticles	60
		2.12.2	Chemical methods	62
		2.12.3	Green synthesis methods	64









	2.12.4	Microwave-assisted green synthesis method	75
2.13	Characte	erization techniques of silver nanoparticles	77
	2.12.1	UV-Visible Spectroscopy	78
	2.12.2	Scanning Electron Microscopy (SEM) & energy-dispersive X-ray spectroscopy (EDX)	79
	2.12.3	X-ray Diffraction (XRD)	80
	2.12.4	Dynamic light scattering (DLS)	81
	2.12.5	Transmission electron microscopy (TEM)	82
2.19	Summar	y for silver nanoparticles	83
2.20	Gap of s	study	85
CHAPTER 3	METH	ODOLOGY	88
3.1 05-4506832 3.2	Introduc pustaka.u Materia		88 n ptbups
3.3	Preparat	ion of Neem extract	91
	3.3.1	Dry process for leaves	91
	3.3.2	Extraction Process	92
	3.3.3	Phytochemical screening of Neem	93
3.4	Synthes	izes of silver nanoparticles (AgNPs)	96
	3.4.1	Green synthesis method	96
	3.4.2	Microwave-assisted green synthesis method	99
3.5	Charact	erization of silver nanoparticles	102
	3.5.1	UV-Visible Spectroscopy (UV-vis)	102
	3.5.2	Fourier-transform infrared spectroscopy (FTIR)	103













Х

dispersive X-ray spectroscopy (EDX)	103
3.5.4 X-RD (X-ray Powder Diffraction)	104
3.5.5 Particle size analysis	104
3.5.6 Transmission electron microscopy (TEM)	105
3.6 Antibacterial activity	105
3.6.1 Preparation of bacterial agar	106
3.6.2 Preparation solutions for antimicrobial test	107
3.7 Preparation of Fish scales Hydroxyapatite/fish scales collagen/ silver nanoparticles composites (FsHA/FsCol/AgNPs)	
3.8 Preparation of Fish Hydroxyapatite/fish collagen/ silver nanoparticles composites Beads	110
05-4506832 3.8.1ka Preparation of FsHA and porous FsHA beads PustakaTBa	inun 1100 ptbupsi
3.8.2 Preparation Beads with collagen and collagen AgNPS	111
3.8.3 Cross-linking of the Beads	112
3.9 Characterizations of Prepared FsHA beads and FsHA composites	115
3.9.1 Fourier transform infrared spectroscopy (FT-IR)	115
3.9.2 Field Emission Scanning Electron Microscopy (FESEM) and EDX FSEM	115
3.9.3 X-ray Powder Diffraction (X-RD)	116
3.10 Antibacterial activity of beads and composites	116
3.10.1 Agar Preparation	116
3.10.2 Antibacterial Test	117















хi

3.11	Bio-compatibility of prepared composites (powder and beads)	118
	3.11.1 Cytotoxicity for FsCol/FsHA/AgNPs composite and beads	118
	3.11.2 Cell attachment for FsCol/FsHA/AgNPs composites	119
3.12	Bio stability and swelling ratios of prepared composites (powder and beads)	120
	3.121 Swelling ratios	120
	3.12.2 Bio stability	120
CHAPTER 4	RESULTS AND DISCUSSION	121
4.1	Neem extraction and phytochemical screening	121
4.2	Comparison between micowave-assisted and green synthesis of silver nanoparticles pustaka.upsi.edu.my Perpustakaan Tuanku Bainun Kampus Sultan Abdul Jalil Shah	123 ptbupsi
	4.2.1 UV-Visible Spectroscopy (UV-vis)	124
	4.2.2 Fourier Transform Infrared spectroscopy (FTIR) Analysis	127
	4.2.3 Scanning electron microscope (SEM)	131
	4.2.4 Energy Dispersive X-Ray Analysis (EDX)	133
	4.2.5 Particle size analysis	136
	4.2.6 XRD (X-ray diffraction) spectroscopy	138
	4.2.7 Mechanism of green synthesis of silver nanoparticles	141
	4.2.8 Summary of results	145
4.3	Effcet of collagen concentration on AgNPs	146
	4.3.1 UV-vis spectroscopy analysis	146
	4.3.2 Fourier-transform infrared spectroscopy (FTIR) analysis	150





















xii

4.3.3	Scanning Electron Microscope (SEM) analysis	152
4.3.4	Energy Dispersive X-Ray Analysis (EDX) analysis	154
4.3.5	Particle size analysis	156
4.3.6	X-Ray Diffraction (XRD) analysis	159
4.3.7	Transmission electron microscopy (TEM) analysis	164
4.3.8	Antibacterial analysis of AgNPs	166
4.3.9	Summary of the Results	169
4.4 Chract	erisation of FsHA/FsCol/AgNPs composites powder	170
4.4.1	Morphology analysis of composites powder	171
4.4.2	Energy Dispersive X-Ray analysis	174
05-4506832 4.4.3 ka	Fourier-transform infrared spectroscopy analysis	177 ptbupsi
4.4.4	X-ray powder diffraction analysis	182
4.4.5	Antibacterial analysis of FsHA/FsCol/AgNPs composite powder	185
4.4.6	Biocompatibility of FsHA/FsCol/AgNPs composite powder	188
	4.4.6.1 Cytotoxicity test	188
	4.4.6.2 Cell attachment study	191
2.4.7	Biostability	195
4.4.8	Swelling ratio	197
4.4.9	Summary of Sesults	198





















xiii

4.5	Synthesis and cheraacteration of FsHA/FsCol/AgNPs composites beads	199
	4.5.1 Morphology and elemental analysis of composites beads	200
	4.5.2 Fourier-transform infrared spectroscopy analysis	215
	4.5.3 X-ray powder diffraction analysis	220
	4.5.4 Antibacterial activates	223
	4.5.5 Bio-compatibility of composites beads	225
	4.5.5.1 Cytotoxicity test of beads	226
	4.5.5.1 Cell attachment of composites beads	227
	4.5.6 Degradation	231
	4.5.7 Swelling ratio	232
05-4506832	4.5.8 ka Summary of results ustakaan Tuanku Bainun Pustaka TBainur	234 ptbups
4.6	The comparison between results that obtained in FsHA/FsCol/AgNPs composites powder and beads	234
CHAPTER 5	CONCLUSIONS AND FUTURE WORKS	236
5.1	Conclusions	236
5.2	Future Works	237
REFERENCES		239
LIST OF PUBLI	ICATIONS	288



















xiv

LIST OF TABLES

T	ables		Page
	2.1	The Incorporated of Col\HA Scaffolds with Materials	35
	2.2	The Collagen and Hydroxyapatite Sources	40
	2.3	Sizes and Shapes with Their Antibacterial Activities of Silver Nanoparticles	47
	2.4	Cytotoxicity studies of silver nanoparticles	57
	2.5	Physical Methods of Silver Nanoparticles with some Applications	62
	2.6	Methods of Silver Nanoparticles with some Applications	63
	2.7	Bacteria Mediated to Synthesis Silver Nanoparticles	66
	2.8 05-4506832 2.9	Fungi Mediated to Synthesis Silver Nanoparticles Perpustakaan Tuanku Bainun Plants Mediated to Synthesis Silver Nanoparticles Pustaka TBainun Plants Mediated to Synthesis Silver Nanoparticles	68 70 ptbups
	3.1	Precursor for Preparation of Different Size of Silver Nanoparticles and its coding	100
	3.2	The Bacteria Strains Used in the Antibacterial Activity	106
	3.3	Formulation of FsHA/ FsCoL/ AgNPs Composites	108
	3.4	Designation of Fish Hydroxyapatite/Collagen Beads Without Silver Nanoparticles	112
	3.5	Designation of Fish Hydroxyapatite/Collagen Beads with Silver Nanoparticles	112
	4.1	Phytochemical Screening of Neem Leaves	122
	4.2	Uv-Vis Absorbance Spectrum of AgC-1 and MAgC-1	126
	4.3	Factional Groups in Fish Collagen and Extract of Neem	129
	4.4	Crystalline particles size of MAgC-1 and AgC-1	140



















χV

	4.5	Uv-Vis Absorbance Spectrum of MAgC-1, MAgC-2, MAgC-3 and MAgC-4	148
	4.6	Factional groups of MAgC-2, MAgC-3 and MAgC-4	151
	4.7	D size of MAgC-2, MAgC-3 and MAgC-4	161
	4.8	The Particle Size and Shape of Ag Nanoparticle Analysed By TEM Technique	164
	4.9	Inhibiting Bacteria from AgC-1 to MAgC-4 against S. <i>aureus</i> and E. <i>Coli</i>	168
	4.10	Factional Groups in FsHA, FsCoL and AgNPs	179
	4.11	Antibacterial activities of control samples against E. coli and S. aureus.	187
	4.12	Antibacterial activities of FsHA/FsCo/AgNPs composites <i>E. coli</i> and <i>S. aureus</i> .	187
)	4.13 05-4506832	Mg-63 Cell Viability Obtained After 24 Hours Exposure to the Test FsHA/FsCol/AgNPs Composites ustakaan Tuanku Bainun Pustaka TBainun Pustaka TBainun	190 ptbupsi
	4.14	Antibacterial activities of BA-1, BB-, BA-3 and BB-3 Beads Against E. coli and S. aureus	225
	4.15	Mg-63 Cell Viability Obtained After 24 Hours Exposure to the Test FsHA/FsCol/AgNPs Composites beads	227





















xvi

LIST OF FIGURES

Fig	ure no		Page
	1.1	The Components of Bone	2
	1.2	Crystal Structure of Hydroxyapatite	3
	1.3	Structure of collagen	5
	2.1	The Proposed Scheme of The Mechanism of Action of Silver Nanoparticles on Bacteria	54
	2.2	Synthesis Methods of Silver Nanoparticles	60
	3.1	Flow Chart of Research Activities a) preparation of Neem extract, b) preparation of AgNPs and c) fabrication of FsHA/Col/AgNPs composites and beads	90
	3.2 05-4506832 3.3	A) Fresh Leaves, B) Dried Leaves of Neem Perpustakaan Tuanku Bainun Preparation of Crude Extract of Neem Abdul Jalil Shah Pustaka TBainun	91 92 ptbups
	3.4	Green Synthesis of Silver Nanoparticles	98
	3.5	Microwave-assisted green synthesis of silver nanoparticle	101
	3.6	Prepration process of fish HA/fish Col/silver nanoparticles composites	109
	3.7	Preparation of 0.5- and 1-MM beads Collagen-Hydroxyapatite-AgNPS	113
	3.8	Preparation of 3 mm beads Collagen-Hydroxyapatite-AgNPS	114
	4.1	Uv-Vis Absorbance Spectrum of a) Neem, b) FsCol, c) AgNO ₃ , d) MAgC-1 and AgC-1	126
	4.2	FTIR spectrum of collagen	128
	4.3	FTIR spectrum of Neem Extract	129













		xvii
4.4	FTIR Spectrum of a) AgC-1 and b) MAgC-2	131
4.5	SEM images of a) AgC-1 and b) MAgC-1	133
4.6	EDX Mapping of a) AgC-1 and b) MAgC-1	135
4.7	Particle Size Analysis of a) AgC-1 and b) MAgC-1	137
4.8	XRD Patterns of a) MAgC-1 and b) AgC-1	139
4.9	Mechanism of silver nanoparticle formation through reduction of silver ion and stabilized by collagen	143
4.10	Mechism of Sample Heating In a) conventional heating and b) microwave heating	145
4.11	Uv-Vis Absorbance Spectrum of Synthesized (a) MAgC-1, (b) MAgC-2, (c) MAgC-3 and (d) MAgC-4	147
4.12	FTIR spectrum of (a) MAgC-1, (b)MAgC-2, (c) MAgC-3 and (d) MAgC-4	150
05-450683 4.13	2 pustaka.upsi.edu.my Perpustakaan Tuanku Bainun Kampus Sultan Abdul Jalil Shah SEM images of a) MAgC-1, b) MAgC-2, c) MAgC -3, and d) MAgC -4	ptbups
4.14	EDX Mapping and Spectrum of (a) MAgC-2, (b) MAgC-3 and (c) MAgC-4	155
4.15	Particle sizes of a) MAgC-2, b) MAgC-3 and c) MAgC-4	157
4.16	XRD Patterns of a) MAgC-2, b) MAgC-3, and c) MAgC-4	160
4.17	Schematic illustration of (a) direct synthesis (in situ) of nanoparticles (NPs) (b) Post synthesis capping by ligand exchange method for the polymer stabilized NPs (Razzaque et al., 2016)	163
4.18	TEM of a) MAgC-2, b) MAgC-3 and c) MAgC-4	165
4.19	Inhibiting Bacteria, a Control samples against a) s. aureus and b) E. Coli	167
4.20	Inhibiting Bacteria from AgC-1 to MAgC-4 against a) S. aureus and b) E. Coli	168













xviii

	4.21	SEM micrograph for a) C-1, b) C-2, c) C-3 and d) C-4 of the FsCol/FsHA/AgNPs composites powder.	172
	4.22	EDX graph for a) C-1, b) C-2, c) C-3 and d) C-4 of FsCol/FsHA/AgNPs	175
	4.23	FTIR analysis of (a) AgNPs, (b) FsCol and (c) FsHA	178
	4.24	Fourier-transform infrared spectroscopy analysis a) C-1, b) C-2, c) C-3, d) C-4, and C-5 FsHA/FsCol/AgNPs composites	180
	4.25	XRD pattern of C-1 (80:20, fish hydroxyapatite/ fish Collagen/silver nanoparticles) composite.	182
	4.26	XRD (X-ray diffraction) analysis of a) C-2 (70:30, FsHA/FsCol/AgNPs), b) C-3 (60:40, FsHA/FsCol/AgNPs c) C-4 (60:40, FsHA/FsCol/AgNPs) and d) C-5 (70:30, FsHA/FsCol/AgNPs) composites.	185
.)	4.27 05-4506832	XRD patterns peaks of a) C-1, b) C-2, c) C-3, d) C-4 and e) C-5 FsHA/FsCol/AgNPs composites Perpustakaan Tuanku Bainun Kampus Sultan Abdul Jalil Shah	184 ptbupsi
	4.28	Antibacterial activities for control samples against a) Staphylococcus aureus and Escherichia coli	185
	4.29	Inhibition of bacteria growth for C-1, C-2, C-3 and C-4 FsCol/FsHA/AgNPs composites against a) <i>Staphylococcus aureus</i> and b) <i>Escherichia coli</i>	186
	4.30	Parantage of MC 62 Call Vishility at Various Concentration of	
		Percentage of MG-63 Cell Viability at Various Concentration of FsHA/FsCol/AgNPs composites	189
	4.31		192
	4.31	FsHA/FsCol/AgNPs composites The Spread of MG-63 cells on the Surface of FsHA/FsCol/AgNPs	
		FsHA/FsCol/AgNPs composites The Spread of MG-63 cells on the Surface of FsHA/FsCol/AgNPs Composites Powder at a) 1, b) 3, c) 7 and d) 14 Days MG-63 attachment on the FsHA/FsCol/AgNPs Composites after a) 1 day, b) 14 days, yellow circle shows the cells attached to the	192
	4.32	FsHA/FsCol/AgNPs composites The Spread of MG-63 cells on the Surface of FsHA/FsCol/AgNPs Composites Powder at a) 1, b) 3, c) 7 and d) 14 Days MG-63 attachment on the FsHA/FsCol/AgNPs Composites after a) 1 day, b) 14 days, yellow circle shows the cells attached to the surface of the composites. Degradation of Ratios of FsHA/FsCol/AgNPs Composite with	192 194





















xix

4.35	SEM Images of a) FsHA//FsCol (without starch) and b) Porous FsHA//FsCol (with starch).	201
4.36	EDX specta of a) FsHA/ FsCol and b) Porous FsHA/ FsCol	203
4.37	SEM Images of (a) FsHA/ FsCol and (b) Porous FsHA/ FsCol	204
4.38	EDX Graph of 1% a) FsHA/ FsCol and b) Porous FsHA/ FsCol	206
4.39	SEM Images of (a) FsHA/ FsCol/AgNPs and (b) Porous FsHA/ FsCol/AgNPs	207
4.40	EDX Graph a) FsHA/ FsCol/AgNPs and b) Porous FsHA/ FsCol/AgNPs	209
4.41	SEM Images of fractured surface (1%) (a) FsHA/ FsCol/AgNPs and (b) Porous FsHA/ FsCol/AgNPs	210
4.42	EDX Graph a) FsHA/ FsCol/AgNPs and b) Porous FsHA/ FsCol/AgNPs	211
4.43 05-4506832 4.44	SEM Images That Expelin Silver Nanoparticles Inside Beads pustaka.upsi.edu.my Perpustakaan Tuanku Bainun Kampus Sultan Abdul Jalil Shah EDX Mapping of Main Elements in a) FsHA/FsCol and b) FsHA/FsCol/AgNPs composites	212 ptbupsi 214
4.45	FTIR spectra of 0.1% FsCol a) FsHA/FsCol and b) Porous FsHA/FsCol composites beads	216
4.46	FTIR spectra of 1% FsCol a) FsHA/FsCol and b) Porous FsHA/FsCol composites beads	217
4.47	FTIR specturm of 0.1% FsCol a) FsHA/FsCol/AgNPs and b) Porous FsHA/FsCol/AgNPs composites beads	218
4.48	FTIR specturm of 1% FsCol a) FsHA/FsCol/AgNPs and b) Porous FsHA/FsCol/AgNPs composites beads	220
4.49	XRD patterns of a) FsHA/FsCol beads and b) Porous FsHA/FsCol composites beads	221
4.50	XRD patterns of a) FsHA/FsCol/AgNPs and b) porous FsHA/FsCol/AgNPs beads	222



















xx

4.51	Inhibition of bacteria growth for FsHA/ FsCol (BA-1), porous FsHA/FsCol A (BB-1) FsHA/ FsCol/ AgNPs (BA-3) and porous FsHA/FsCol/ AgNPs (BB-3) beads against a) <i>Escherichia coli</i> and b) <i>Staphylococcus aureus</i> .	224
4.52	Percentage of MG-63 Cell Viability at Various Concentration of FsHA/FsCol/AgNPs composites beads	226
4.53	The Spread of MG-63 cells on the Surface of FsHA/FsCOI/AgNPs Composites (beads) at a) 1, b) 3, c) 7 and d) 14 Days	228
4.54	MG-63 attachment on the FsHA/FsCol/AgNPs Composites beads for a) 1 day and b) 14 days. orange circles show the attached for cells in the beads	230
4.55	Degradation of Ratios of FsHA/FsCol (BA-1), porous FsHA/FsCol (BB-1), FsHA/FsCol/ AgNPs (BA-3) and porous FsHA/FsCol/ AgNPs (BB-3) bead	231
4.56	Swelling rate of FsHA/ FsCol (BA-1), porous FsHA/FsCol (BB-1), FsHA/ FsCol/ AgNPs (BA-3) and porous FsHA/ FsCol/ AgNPs (BB-3) beads	232
) 05-4506832	Paraustakaan Tuanku Pajaun	



















xxi

LIST OF ABBREVIATIONS

AB assay Alamar Blue Assay

AFM Atomic Force Microscopy

Silver Ag

AgNO₃ Silver Nitrate

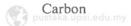
AgNPs Silver Nanoparticles

Al Aluminium

BG Bioactive Glass

BMP-2 Morphogenetic Protein-2

05-4506832





PustakaTBainun



Calcium

Cap Calcium Phosphate

CHC₁₃ Chloroform

Chlorine CI

CPC Calcium Phosphate Cement

CS Chitosan

Cu Copper

DAPI 4',6-Diamidino-2-Phenylindole

DCPA Dicalcium Phosphate Anhydrous

DCPD Dicalcium Phosphate Dehydrate

Dehydrothermal DHT





















xxii

DLS Dynamic Light Scattering

DMEM Dulbecco'S Modified Eagle'S Medium

DMF N, N-Dimethylformamide

DW Distilled Water

E. coli Escherichia Coli

EDTA Ethylenediaminetetraacetic Acid

EDX Energy Dispersive X-Ray Analysis

FAK Focal Adhesion Kinase

Fe Iron

FGF-2 Growth Factor Of Fibroblast

FsCol Fish Scals Collagen

FsHA 4506832 Fish Scals Haydroxyapitite Sultan Abdul Jalil Shah

FTIR Fourier-Transform Infrared Spectroscopy

FWHM Full-Width At Half-Maxima

GS Green Synthesis

HCl Hydrochloric Acid

HMD Hyaline Membrane Diseases

LAT Laser Ablation Technique

MAGS Microwave-Assisted Green Synthesis

Mg Magnesium

MSCs Mesenchymal Stem Cells

N Nitrogen

NA Nutrient Agar









PustakaTBainun













xxiii

Na Sodium

NaBH₄ Sodium Borohydride

NCPs Non-Collagenous Proteins

NE Neem Extract

NGF-β Nerve Growth Factor

NPs Nanoparticles

O Oxygen

OC Osteochondral

P Phosphorus

PBS Phosphate Buffer Saline

PCL Polycaprolactone

PLA⁰⁵⁻⁴⁵⁰⁶⁸³²



Poly (Lactic Acid)

Perpustakaan Tuanku Bainun Kampus Sultan Abdul Jalil Shah



PustakaTBainun



ptbups

PLCL Poly (Lactide-Co-Ecaprolactone)

PLGA Poly-(Lactide-Co-Glycolide

PLLA Poly (L-Lactic Acid)

PMMA Poly (Methyl Methacrylate

Poly-HEMA Poly-Hydroxyethyl Methacrylate

PVA Poly (Vinyl Alcohol)

PVP Polyvinylpyrrolidone

ROS Reactive Oxygen Species

S. aureus Staphylococcus Aureus

SBF Simulated Body Fluid

SEM Scanning Electron Microscope





















xxiv

Si Silicon

Surface Plasmon Resonance SPR

TEM Transmission Electron Microscopy

TTCP Tetracalcium Phosphate

Urinary Tract Infections **UTIs**

Ultraviolet-Visible Spectroscopy **UV-Vis**

X-Ray Photoelectron Spectroscopy **XPS**

X-Ray Diffraction Analysis **XRD**

β Beta

β ΤСР B-Tricalcium Phosphate

X-Ray Radiation





























CHAPTER 1

INTRODUCTION











1.1 Background of study

Bone is considered the core unit that forms the human body's skeleton, consisting primarily of hydroxyapatite (HA) and collagen (Bandyopadhyay, 2008). Bone consists of four main components: HA minerals, type-I collagen, water, and non-collagenous proteins (NCPs). By weight, bone is ~70 wt % minerals, ~20 wt % organics, and ~10 wt % water. The structure of natural bone is shown in figure 1.1. Natural bone is a composite material comprised of organic and inorganic elements (Andric et al., 2011). The organic materials are mainly collagen fibers containing tropocollagen, which make up most of the organic constituent of bone, and provide strength to the bone (Basha et al., 2015). The inorganic materials are mainly calcium (Ca) and phosphorus (P) in the form



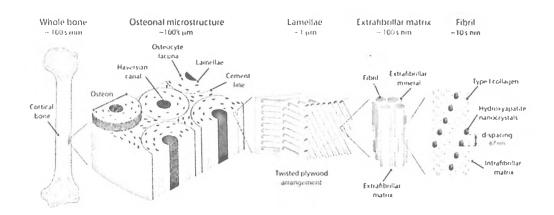








of hydroxyapatite (HA). Kamitakahara et al., (2007), howver reported bone is a composite of 70 mass% inorganic component, i.e., hydroxyapatite, and 30 mass% organic components, i.e., collagen (Kamitakahara et al., 2007).



05-4506832 pustaka upst.edu.my Perpustakaan Tuanku Bainun Pustaka TBainun Figure 1.1. The Components of Bone (https://www.nature.com/articles/boneres201759)

Bone defects may occur for several reasons, such as injuries, illnesses, surgical interventions, and accidents, some of which may heal on their own. However, bone defects greater than 1/3 an inch (\approx 8 mm) cannot be healed on their own (Gupta, Thussbas, Koch, & Seebauer, 2018). Therefore, bone substitutes were used to fill up and enhance bone defects to allow for the rapid healing process (Qian et al., 2018). These substitutes provide structural and mechanical support to enhance bone tissue formation or fill gaps to facilitate the healing of bone tissue. Bone substitutes have been widely used in plastic surgery, oral, maxillofacial, dental, and orthopedic surgery, making it one of the most implanted tissues in the medical field (Fliefel, Ehrenfeld, & Otto, 2018).











HA is an inorganic compound with the chemical structure Ca₁₀(PO4)₆ (OH)₂. It is a type of biomaterial widely used for orthopedics and dental due to its bioactive properties and osteoconductive properties (Habibah & Salisbury, 2018). The crystal structure of HA is shown in Figure 1.2. HA has been used in many composites to enhance the healing of bone defects, such as HA-poly(lactic acid) as an artificial bone filler (Higashi et al., 1986), HA-polyamide as an osteoconductive agent (Lu et al., 2016), nHA (Nano-HA)chitosan for orthopedic bone (Zhang et al., 2012), nHA/gelatin for pharmaceutical and cosmetic purposes (Mohamed, Beherei, & El-Rashidy, 2014).

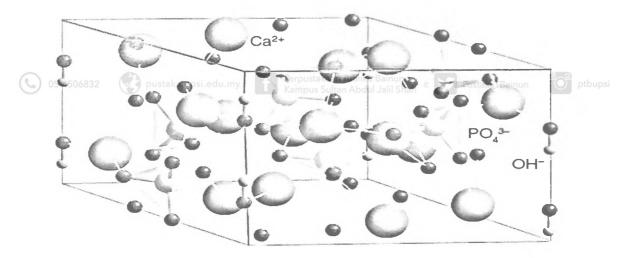


Figure 1.2. Crystal Structure of Hydroxyapatite (https://www.c-hemtube3d.com/ss hydroxyapatite /)

Most of the HA used in commercial products are derived through chemical synthesis. However, the high price of chemically-synthesized HA has motivated researchers to find a solution for other sources of HA. One of the alternatives is natural HA from animal bones (Sobczak-Kupiec & Wzorek, 2012). However, the recent problem of animal



















diseases such as hyaline membrane disease (HMD) and mad cow disease renders animal-based HA not a good alternative of HA. As an alternative, HA from fish scale was studied and found to be the best substitute compared to synthetic HA or animal-based HA (Zainol et al., 2012).

Collagen, a natural polymer that can be isolated from different animal parts such as ligaments, tendons, skin, and bone, is used as a biomaterial due to its biocompatibility (Jia et al., 2013). Collagen has been proven to enhance cell growth and has excellent biological properties (Takitoh et al., 2015; Maslennikova et al., 2015). The structure of collagen is shown in Figure 1.3. Many studies reported the development of highly-modified collagen through the incorporation of different materials, and its use to enhance the healing of bone defects, such as collagen-β-tricalcium phosphate to improve osteoconductivity (Yunoki et al., 2006), collagen-calcium phosphate to improve the biodegradability materials (Du et al., 2000), collagen-poly(lactic-co-glycolic acid) for the healing of cartilage defect (Dean et al., 2003), and collagen-chitosan-glycerin as biostability and biocompatibility materials (Ullah, Zainol, Chowdhury, & Fauzi, 2018).











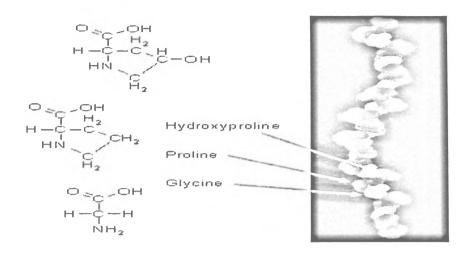


Figure 1.3. Structure of collagen (Silvipriya et al., 2015)

Among several biomaterials investigated to treat bone defects, the composites material exhibited the best effect and can be utilized clinically. HA/collagen composites have been prepared using various preparation techniques and have been used in many bone defects as a biomaterial for bone tissue engineering. Various sizes (nano-size and micro-size) of the Col (collagen)/HA (hydroxyapatite) composites were incorporated with different materials such as poly (l-lactic acid; PLLA) (Liao & Cui, 2004), poly(vinyl alcohol) (PVA); (Song et al., 2012), chitosan (Zhang, Tang, Zhang, Xu, & Wang, 2009), and metals such as Fe (Tampieri et al., 2014), to develop the properties of the scaffolds that exhibit high compatibility and bio-degradable properties for bone repair. The collagen/HA (Col/HA) composites used in many medical applications were fabricated from various sources. However, there was no report on the application of fish scales collagen and fish scales hydroxyapatite as Col/HA composites.



















Col/HA composites were formulated in different forms such as powder, paste, gel, sponge, and beads. Currently, Morra et al., (2015), reported the preparation of hydroxyapatite (HA)/β TCP (β-tricalcium phosphate)/collagen beads as bone fillers. They placed the HA beads in the collagen solution to allow even distribution of collagen in the porous HA beads. The HA/collagen mixture was then characterized using XPS and FTIR to confirm the structure of the beads. The HA was derived from chemical synthesis, whereas the collagen was extracted from the animal.

The incorporation of HA and collagen is possible due to the biocompatibility of collagen and high mechanical properties of the HA, and it is widely utilized as the biomaterial to enhance the healing of bone defect, as well as a replacement material for bone defects. Nonetheless, despite its advantages, several issues of collagen/HA composite concern the public. Tampieri et al. (2014) reported the successful development of collagen/HA composite that showed excellent bioactivity properties, which lead to its use as a bone filler for the defect bones. However, most of the collagen used in the market is extracted from animals. Thus, halal issues became a significant concern for Muslim users.

Another issue of concern for the consumers is the failure of the implant or foreign body used in orthopedic surgery due to bacterial infections (Ahmed et al., 2016). The adhesion of bacteria to an implant occurs in the form of a biofilm, rendering the bacteria extremely resistant to the host's defense mechanisms and antimicrobials (Dastgheyb et al., 2015). Nonetheless, despite the recognized need for implant-related infection





















containment and the demonstrated efficacy of some antibacterial coatings, only a few technologies are currently available in orthopedics and trauma (Berend et al., 2013). While some potentially effective solutions have been identified as not suitable for orthopedic implants due to their cytotoxicity, immunoreactivity, or interference with bone healing and osteointegration, those successfully tested in vitro and in vivo may still not be able to achieve large-scale clinical application due to the biotechnological, economy, and regulatory issues (Kurtz et al., 2007). Silver nanoparticles used as agents to inhibit bacterial growth (Chaloupka, Malam, & Seifalian, 2010) have also shown their ability to function as an antibacterial agent when incorporated with composite scaffolds for bone defects (Saravanan et al., 2011). The use of silver nanoparticles (AgNPs) synthesized by chemical, physical, and biological methods helped minimize bacterial infections after bone defect filling, as many unsuccessful orthopedic surgeries were due to bacterial publications. infections (Ahmed et al., 2016). Due to the environmental risks of nanomaterials, which were created by chemical and physical processes, several techniques were developed to solve the problem and to create environmentally friendly, non-toxic materials (Ahmed et al., 2016).

The green synthesis of AgNPs have been conducted by utilizing several plants extract such as Coffea arabica (Dhand et al., 2016), Euphorbia tirucalli (Kalaiselvi, Mohankumar Shanmugam, Nivitha, & Sundararaj, 2019), Annona squamosal (Ruddaraju, Pallela, Pammi, Padavala, & Kolapalli, 2019), Prosopis juliflora (Arya et al., 2018). Nanoparticles have been investigated as an excellent antimicrobial agent and applied for many applications such as arthroplasty (Marin et al., 2015), catheters (Rafique, Sadaf,





















Rafique, & Tahir, 2017), stainless steel materials (Mochochoko, Oluwafemi, Jumbam, & Songca, 2013), dental materials (Emmanuel et al., 2015), and human skin (AlSalhi et al., 2016). Neem is one of the Melia genera belonging to the Meliaceae family, which is widely distributed in India and Malaysia. The Neem has been extracted and used to produce nanosilver, which has shown excellent results by using silver nitrate for anticancer activity (Kathiravan et al., 2014).

In this study, the microwave-assisted green synthesis of silver nanoparticles was conducted using neem extract and collagen as a reducing and stabilizing agent, respectively. The Ag-stabilized collagen was used as an enhancement to infiltrate into the HA beads to form FsHA/FsCol/AgNPs composite. The beads of FsHA/FsCol/AgNPs have been formulated and characterized for bone fillers applications.

1.2 Problem statements

The need for the grafts of bones or substitutes increased with the increasing need for suitable materials to be used as substitutes in many cases, such as the treatment of fractures revision hip arthroplasty, spine fusion, and tumors in addition to reconstructive surgery. This situation renders researchers look for vital materials that can be utilized to perform this purpose (Pedersen et al., 2005). Several requirements apply to bone fillers and any other implantable biomaterials, i.e., they must be non-toxic and biocompatible,





















)

do not initiate an adverse inflammatory reaction, and must be commercially viable to manufacture and cost-effective for the surgeon to use.

The high cost of chemicals used in the synthesis of HA has motivated researchers to find a solution for another source of HA. One of the alternatives is natural HA from animal bones (Sobczak-Kupiec & Wzorek, 2012). However, animal diseases such as hyaline membrane disease (HMD) and mad cow disease poses a threat to the production of HA from animal bones, rendering animal-based HA not a good alternative of HA. Thus, the fish scale HA was studied and was found to be the best alternative compared to synthetic HA or animal-based HA (Zainol et al., 2012). Fish scales consist of 50 wt% collagen and 50 wt% HA and could be extracted from fish scales using a simple extraction method (Zainol et al., 2012). FsHA has been used in many biological and medical applications due to osteoinductive and osteoconductive properties. Other characteristics that make HA attractive for use as bone replacement are that it is a significant component of bones and hard tissues, is non-toxic and nonimmunogenic, has the mechanical strength and surface properties that are suitable for bone tissue regeneration (Pon-On et al., 2016). However, the report regarding the potential use of fish scales as HA/collagen composites as bone filler is lacking.

Despite the increasing utilization of implanted biomaterials, their long-term durability is not guaranteed, besides the risk of infection that dictates for early failure in orthopedics and trauma. The economic and social costs of implant-related infections are significant. Silver nanoparticles can reduce the potential of bacterial infections from





















implant materials (Moriarty et al., 2016). The antibacterial activity of silver nanoparticles is well known, mostly dependent on dissolved cations' ability to interfere with bacterial cell membrane permeability and cellular metabolism (Schmidt-Braekling et al., 2017). However, a few implant designs are incorporated with silver due to the cost of the technology. Therefore, to solve this problem, the current study utilized low-cost silver nanoparticles prepared from natural materials.

Based on previous studies, no research was reported on the incorporation of silver nanoparticles in HA/collagen composites. Thus, a novel fish scales HA/collagen/silver composite was developed for cheaper and safer bone filler materials in this study.











1.3 Research objectives

- 1. To synthesis nanosilver particles using microwave-assisted green synthesis with neem (M. dubia) as a reducing agent and collagen as a stabilizer.
- 2. To formulate and characterize fish scales HA/collagen/silver nanoparticles (FsHA/FsCol/AgNPs) composites for bone filler applications.
- 3. To investigate the antibacterial activities of FsHA/FsCol/AgNPs composites against pathogenic Escherichia coli and Staphylococcus aureus.
- 4. To evaluate the biostability and biological properties of FsHA/FsCol/AgNPs composites.





















1.4 Significant of the study

The use of biomaterials to fabricate a suitable substance for an implant in many applications such as bone graft, drug delivery, bone fillers, and tissue engineering of the bone has been one of the significant challenges for many years. HA is one of the materials that have been reported to have excellent properties as a biomaterial, such as biocompatibility, non-inflammatory, non-toxic, osteoconductivity, and none-immunogenicity. The fabrication of collagen with HA is suitable as bone fillers, as well as bone graft and other biomaterials field due to its excellent mechanical properties, biological activity, and others. This study aimed to improve the biological properties of fish scales HA/fish collagen through incorporation with silver nanoparticles, which exhibits antimicrobial activity that allow the beads to be successfully used in bone graft and bone fillers surgery. This preparation could overcome the problems due to bacteria infection.

This study successfully produced FsHA/FsCol/AgNPs composite that possesses biological properties and mechanical properties suitable for bone fillers. The product is relatively cheap and affordable for the public. Due to low cost of raw materials i.e fish scales. Furthermore, the material used is halal and suitable for Muslims around the world.









