



ISOLATION OF ALKALOIDS FROM *Alseodaphne pendulifolia* Gamb. (LAURACEAE) AND THEIR BIOLOGICAL ACTIVITIES



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MUHAMMAD HAFIZ HUSNA BIN HASNAN



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UNIVERSITI PENDIDIKAN SULTAN IDRIS**

2014



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This declaration is made on the 8th day of AUGUST 2014.

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I, MUHAMMAD HAFIZ HUSNA BIN HASNAN, M20102001203, F3MT (PLEASE INDICATE STUDENT'S NAME, MATRIC NO. AND FACULTY) hereby declare that the work entitled ISOLATION OF ALKALOIDS FROM ALSEODAPHNE PENDULIFOLIA GAMB. (LAURACEAE) AND THEIR BIOLOGICAL ACTIVITIES is my original work. I have not copied from any other students' work or from any other sources except where due reference or acknowledgement is made explicitly in the text, nor has any part been written for me by another person.

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ABSTRACT

The purpose of this study is to extract, isolate and elucidate the structure of alkaloid constituents from barks of *Alseodaphne pendulifolia* Gamb. (Lauraceae) and to determine the biological activities (antiplasmodial, cytotoxicity, antibacterial and antioxidant) against the isolated alkaloids. The samples were collected from Sg. Temau Forest Reserve, Kuala Lipis, Pahang and Sg. Tekam Forest Reserve, Jerantut, Pahang, Malaysia. The isolation and purification of alkaloids were done using various chromatographic techniques including chromatotron, column, preparative thin layer and thin layer chromatography. The structure elucidations of the isolated alkaloids were established by NMR spectral data using various 1D-NMR (^1H , ^{13}C , and DEPT) and 2D-NMR (COSY, HMQC/HSQC, HMBC, H2BC, HSQC-TOCSY and NOESY) techniques, aided via UV, IR, MS data and also by comparison with data from the literature. Further isolation process of the plants yielded four types of isoquinoline alkaloid; aporphine, aporphine *N*-oxide, oxoaporphine and bisbenzylisoquinoline. A new aporphine *N*-oxide; norisocorydine *N*-oxide **166** with four aporphines; corydine **124**, *N*-methylhernagine **169**, oxohernagine **122** and hernagine **123**, were successfully isolated from samples that collected from Sg. Temau Reserve Forest. Samples from Sg. Tekam Reserve Forest, yielded four aporphines; corydine **124**, *N*-methylhernagine **169**, hernagine **123** and hernovine **170**, an oxoaporphine; oxohernagine **122** was isolated together with a bisbenzylisoquinoline; costaricine **126**. Antiplasmodial assay against *Plasmodium falciparum* on the bark crude extract exhibited active antiplasmodial activity with IC_{50} value 0.804 $\mu\text{g}/\text{ml}$, while the isolated pure compounds corydine **124**, oxohernagine **122** and hernagine **123** showed the active antimalaria activity IC_{50} values at 1.184, 2.189 and 3.494 μM , respectively. The new discovered compound, norisocorydine *N*-oxide showed a moderate effect on cytotoxic assay using leukaemia cell lines HL-60 with IC_{50} value 11 $\mu\text{g}/\text{ml}$. Norisocorydine *N*-oxide **166**, corydine **124**, *N*-methylhernagine **169**, oxohernagine **122** and hernagine **123** exhibited very weak activity with IC_{50} values more than 500 $\mu\text{g}/\text{mL}$ against free radical scavenging effect of antioxidant DPPH assay and were not active against gram positive bacteria; *Bacillus subtilis* (B145). This is the first report for phytochemical and biological activity studies from barks of *Alseodaphne pendulifolia* Gamb., which has potential in determination of other various chemical constituents for new drug discovery.





PEMENCILAN ALKALOID DARIPADA *Alseodaphne pendulifolia* Gamb. (LAURACEAE) DAN AKTIVITI-AKTIVITI BIOLOGI

ABSTRAK

Kajian ini bertujuan untuk mengekstrak, memencil dan menentukan struktur sebatian alkaloid daripada kulit batang *Alseodaphne pendulifolia* Gamb. (Lauraceae) dan menentukan aktiviti biologi (antiplasmodial, sitotoksiti, antibakteria, dan antioksidan) terhadap sebatian alkaloid yang telah dipencilkan. Sampel diperoleh dari Hutan Simpan Sg. Temau, Kuala Lipis, Pahang dan Hutan Simpan Sg. Tekam, Jerantut, Pahang, Malaysia. Proses pemencilan dan penulenan alkaloid menggunakan pelbagai kaedah kromatografi termasuk kromatotron, kromatografi turus, kromatografi plat dan kromatografi lapisan nipis. Pengenalpastian struktur alkaloid yang telah dipencil dan ditentukan melalui data spektrum NMR menggunakan pelbagai teknik 1D-NMR (^1H , ^{13}C , dan DEPT) dan 2D-NMR (COSY, HMQC/HSQC, HMBC, H2BC, HSQC-TOCSY dan NOESY), serta disokong oleh data daripada UV, IR, MS, dan juga melalui perbandingan data kajian literatur. Proses pemencilan sebatian kimia terhadap tumbuhan ini telah berjaya memperoleh empat alkaloid jenis isokuinolina; aporfina, aporfina *N*-oksida, oxoaporfina dan bisbenzilisokuinolina. Sebatian alkaloid aporfina *N*-oksida yang baru ditemui; norisokoridina *N*-oksida **166** dan empat lagi aporfina; koridina **124**, *N*-metilhernagina **169**, oxohernagina **122** dan hernagina **123** telah berjaya dipencilkan daripada sampel yang diperoleh dari Hutan Simpan Sg. Temau. Bagi sampel yang diperolehi dari Hutan Simpan Sg. Tekam, empat aporfina telah berjaya dipencilkan; koridina **124**, *N*-metilhernagina **169**, hernagina **123** dan hernovina **170**, satu oxoaporfina; oxohernagina **122** juga dipencilkan bersama-sama dengan satu sebatian bisbenzilisokuinolina; kostaricina **126**. Ujian antiplasmodial *Plasmodium falciparum* ke atas ekstrak mentah menunjukkan aktiviti antimalaria yang aktif dengan nilai IC_{50} sebanyak $0.804 \mu\text{g/ml}$, sementara untuk sebatian yang dipencilkan koridina **124**, oxohernagina **122** dan hernagina **123** menunjukkan aktiviti yang aktif iaitu nilai IC_{50} masing-masing sebanyak 1.184 , 2.189 and $3.494 \mu\text{M}$. Ujian ketoksikdan terhadap sel leukimia HL-60, mendapati sebatian baru; norisokoridina *N*-oxide **166** menunjukkan kesan sederhana dengan nilai IC_{50} $11 \mu\text{g/ml}$. Sebatian norisokoridina *N*-oksida **166**, koridina **124**, *N*-metilhernagina **169**, oxohernagina **122** dan hernagina **123** menunjukkan aktiviti yang sangat lemah dengan nilai IC_{50} melebihi $500 \mu\text{g/mL}$ terhadap kesan penjerapan radikal bebas dalam ujian antioksida DPPH dan juga menunjukkan kesan tidak aktif melawan bakteria gram positif: *Bacillus subtilis* (B 145). Ini merupakan laporan pertama mengenai kajian fitokimia dan aktiviti-aktiviti biologi terhadap kulit batang *Alseodaphne pendulifolia* Gamb. yang mana berpotensi untuk menemukan pelbagai jenis sebatian kimia yang lain untuk penghasilan ubat-ubatan baru.



CONTENTS

	PAGE
DECLARATION	ii
ACKNOWLEDGEMENT	iii
ABSTRACT	iv
ABSTRAK	v
CONTENTS	vi
LIST OF TABLES	ix
LIST OF FIGURES	xi
LIST OF SCHEMES	xiv
LIST OF CHEMICAL COMPOUND STRUCTURE	xv
ABBREVIATIONS	xix

CHAPTER 1 INTRODUCTION	1
1.1 General	1
1.2 Problem Statements	4
1.3 Objectives of Study	7
1.4 Family of Lauraceae	8
1.5 Genus of <i>Alseodaphne</i>	12
1.6 Species of <i>Alseodaphne pendulifolia</i> Gamb.	17
 CHAPTER 2 GENERAL CHEMICAL ASPECTS	 19
2.1 General	19
2.2 Alkaloid	21

CONTENTS (cont.)

2.3	Classifications of Alkaloids	24
2.3.1	True Alkaloids	24
2.3.2	Protoalkaloids	25
2.3.3	Pseudoalkaloids	25
2.4	Isoquinoline Alkaloids	28
2.5	Aporphine Alkaloid	31
2.6	Bisbenzylisoquinoline Alkaloid	36
2.7	Chemical Constituent from Genus of Alseodaphne	43

CHAPTER 3 MATERIAL AND EXPERIMENTAL 58

3.1	General	58
3.2	Isolation and Structure Determination of Alkaloids	58
3.2.1	Instruments	59
3.2.2	Reagents	60
3.2.3	Plants Materials	61
3.2.4	Alkaloids Plants Screening	61
3.2.5	Extraction of Plant Materials	62
3.2.6	Separation and Purification of Alkaloids	63
3.2.7	Physical and Spectral Data of Isolated Alkaloids	66
3.3	Biological Activity Assay	73
3.3.1	<i>In-vitro</i> Antiplasmodial Assay	73
3.3.2	Cell Culture and MTT Cytotoxicity Assay	74

CONTENTS (cont.)

3.3.3	Bacterial Culture and Disc Diffusion Assay	75
3.3.4	DPPH Free Radical Scavenging Activity Assay	76
CHAPTER 4	RESULTS AND DISCUSSION	78
4.1	Isolation of Alkaloids from Barks of <i>Alseodaphne pendulifolia</i> Gamb.	78
4.2	Structure Elucidation of Isolated Alkaloid from Barks of <i>Alseodaphne pendulifolia</i> Gamb.	82
4.2.1	APB 1, Norisocorydine <i>N</i> -oxide 166	82
4.2.2	APB 2, Corydine 124	93
4.2.3	APB 3, <i>N</i> -methylhernagine 169	101
4.2.4	APB 4, Oxohernagine 122	109
4.2.5	APB 5, Hernagine 123	116
4.2.6	APB 6, Hernovine 170	123
4.2.7	APB 7, Costaricine 126	131
4.3	Biological Activity of Alkaloid Compounds from Barks of <i>Alseodaphne pendulifolia</i>	139
CHAPTER 5	CONCLUSION	142
REFERENCES		144
APPENDIXES 1		158
	Articles in Proceeding/Presented at Conferences/Seminar	
APPENDIXES 2		181
	Antiplasmodial Results Details	

LIST OF TABLES

Table	Title	Page
1.1.1	Specific Plants Parts in Certain Plant Species Used in Phytotherapy.	4
1.3.1	Uses of Species from Family of Lauraceae	11
1.4.1	List of Species in Genus of <i>Alseodaphne</i> (Lauraceae)	14
2.4.1	Examples of Alkaloids from Isoquinoline Subgroups and Their Source.	29
2.6.1	Groups and Subgroups for Classification of Bisbenzylisoquinoline Alkaloids and Their Examples	39
2.8.1	Chemical Constituents Isolated from Genus of <i>Alseodaphne</i>	43
3.2.4.1	Indicator for Mayer's Test on Alkaloid	62
3.2.5.1	Yield of Crude Alkaloids Extracted from Barks of <i>Alseodaphne pendulifolia</i> Gamb. (Lauraceae)	63
4.2.1.1	¹ H NMR Spectral Comparison of APB 1 with Isocorydine 94 and Norisocorydine 95	86
4.2.1.2	¹ H NMR Spectral Comparison between Corydine N-oxide 167 toward Corydine 124 (Ferreira et al., 2010)	86
4.2.1.3	1D (¹ H, ¹³ C and DEPT-135), 2D (COSY, HMQC and HMBC) NMR Spectral Data of APB 1	87
4.2.2.1	1D (¹ H and ¹³ C) and 2D (COSY, HMQC and HMBC) NMR Spectral Data of APB 2	99
4.2.2.2	¹ H And ¹³ C NMR Spectral Data Comparison of APB 2, with Corydine 124 from Literature	100
4.2.3.1	1D (¹ H, ¹³ C and DEPT-135) and 2D (COSY, HMQC and HMBC) NMR Spectral Data of APB 3	107
4.2.3.2	¹ H NMR Spectral Data Comparison of APB 3, with N-methylhernagine 169 from Literature (Guinaudeau et al., 1983)	108
4.2.4.1	1D (¹ H, ¹³ C And DEPT-135) And 2D (COSY, HMQC and HMBC) NMR Spectral Data of APB 4	114



LIST OF TABLES (cont.)

4.2.4.2	¹ H NMR Spectral Data Comparison of APB 4, with Oxohernagine 122 from Literature (Chen Et Al., 1996)	115
4.2.5.1	1D (¹ H, ¹³ C and DEPT-135) and 2D (COSY, HMQC and HMBC) NMR Spectral Data of APB 5	119
4.2.5.2	¹³ C NMR Spectral Data Comparison of APB 5, with Hernagine 123 in Literature Data (Phan et al., 1994)	122
4.2.6.1	1D (¹ H, ¹³ C and DEPT-135) and 2D (COSY, HMQC and HMBC) NMR Spectral Data of APB 6	129
4.2.6.2	¹³ C NMR Spectral Data Comparison of APB 6, with Hernovine 170 in Literature Data (Phan et al., 1994)	130
4.2.7.1	1D (¹ H, ¹³ C and DEPT-135) and 2D (COSY, HMQC and HMBC) NMR Spectral Data of APB 7	137
4.2.5.2	¹³ C NMR Spectral Data Comparison of APB 7, with Costaricine 126 from Literature (Phan et al., 1994)	138
4.3.1	Biological Activities (<i>In-Vitro</i> Antiplasmodial HRP2, Cytotoxic, Antibacterial and Antioxidant) of Alkaloids Against Alkaloids Crude Extract and Isolated Aporphine Alkaloids from Barks of <i>Alseodaphne pendulifolia</i> Gamb.	140





LIST OF FIGURES

Figures	Title	Page
1.5.1	Fruits of <i>Alseodaphne pendulifolia</i>	18
1.5.2	Barks of <i>Alseodaphne pendulifolia</i>	18
1.5.3	Leaves of <i>Alseodaphne pendulifolia</i>	18
2.5.1	Numbering of Aporphine Skeleton	33
2.5.2	Configuration of Aporphine; Left – A Configuration of Aporphine; Right – B Configuration of Aporphine	33
2.6.1	Numbering system of Bisbenzylisoquinoline Alkaloids	37
4.2.1.1	¹ H NMR Spectrum of APB 1	85
4.2.1.2	¹³ C NMR Spectrum of APB 1	88
4.2.1.3	DEPT-135 NMR Spectrum of APB 1	88
4.2.1.4	COSY NMR Spectrum of APB 1	89
4.2.1.5	HSQC NMR Spectrum of APB 1	89
4.2.1.6	HMBC NMR Spectrum of APB 1	90
4.2.1.7	COSY and ¹ H- ¹³ C Coupling Pattern Observed in HMBC Spectrum of APB 1	90
4.2.1.8	NOESY NMR Spectrum of APB 1	91
4.2.1.9	H2BC NMR Spectrum of APB 1	91
4.2.1.10	HSQC-TOCSY NMR Spectrum of APB 1	92
4.2.2.1	¹ H NMR Spectrum of APB 2	96
4.2.2.2	¹³ C NMR Spectrum of APB 2	97
4.2.2.3	COSY NMR Spectrum of APB 2	97
4.2.2.4	HMQC NMR Spectrum of APB 2	98



**LIST OF FIGURES (cont.)**

4.2.2.5	HMBC NMR Spectrum of APB 2	98
4.2.2.6	COSY and ^1H - ^{13}C Coupling Pattern Observed in HMBC NMR Spectrum of APB 2	99
4.2.3.1	^1H NMR Spectrum of APB 3	103
4.2.3.2	^{13}C NMR Spectrum of APB 3	104
4.2.3.3	DEPT-135 NMR Spectrum of APB 3	104
4.2.3.4	COSY NMR Spectrum of APB 3	105
4.2.3.5	HSQC NMR Spectrum of APB 3	105
4.2.3.6	HMBC NMR Spectrum of APB 3	106
4.2.3.7	COSY and ^1H - ^{13}C Coupling Pattern Observed in HMBC NMR Spectrum of APB 3	106
4.2.4.1	^1H NMR Spectrum of APB 4	111
4.2.4.2	^{13}C NMR Spectrum of APB 4	111
4.2.4.3	DEPT-135 NMR Spectrum of APB 4	112
4.2.4.4	COSY NMR Spectrum of APB 4	112
4.2.4.5	HSQC NMR Spectrum of APB 4	113
4.2.4.6	HMBC NMR Spectrum of APB 4	113
4.2.4.7	COSY and ^1H - ^{13}C Coupling Pattern Observed in HMBC NMR Spectrum of APB 4	114
4.2.5.1	^1H NMR Spectrum of APB 5	118
4.2.5.2	^{13}C NMR Spectrum of APB 5	118
4.2.5.3	DEPT-135 NMR Spectrum of APB 5	120
4.2.5.4	HMQC NMR Spectrum of APB 5	120
4.2.5.5	COSY NMR Spectrum of APB 5	121



**LIST OF FIGURES (cont.)**

4.2.5.6	HMBC NMR Spectrum of APB 5	121
4.2.5.7	COSY and ^1H - ^{13}C Coupling Pattern Observed in HMBC NMR Spectrum of APB 5	122
4.2.6.1	^1H NMR Spectrum of APB 6	125
4.2.6.2	^{13}C NMR Spectrum of APB 6	126
4.2.6.3	Dept-135 NMR Spectrum of APB 6	126
4.2.6.4	COSY NMR Spectrum of APB 6	127
4.2.6.5	HMQC NMR Spectrum of APB 6	127
4.2.6.6	HMBC NMR Spectrum of APB 6	128
4.2.6.7	COSY and ^1H - ^{13}C Coupling Pattern Observed in HMBC NMR Spectrum of APB 6	128
4.2.7.1	^1H NMR Spectrum of APB 7	133
4.2.7.2	^{13}C NMR Spectrum of APB 7	134
4.2.7.3	COSY NMR Spectrum of APB 7	134
4.2.7.4	HMQC NMR Spectrum of APB 7	135
4.2.7.5	HMBC NMR Spectrum of APB 7	135
4.2.7.6	Dept-135 NMR Spectrum of APB 7	136
4.3.7.7	COSY and ^1H - ^{13}C Coupling Pattern Observed in HMBC NMR Spectrum of APB 7	136





LIST OF SCHEMES

Scheme	Title	Page
2.5.1	The Various Groups of Benzyloquinoline Derived from (S)-Norcoclaaurine 59	34
2.5.2	Scheme: The Biogenetic Pathway from 49 to Corytuberine 60 , Isoboldine 61 , Glucine 62 and Bulbocapnine 63	35
2.6.1	The Biogenetic Pathway of Berbamunine	38
4.1.1	Isolation of Alkaloid from the Barks of <i>Alseodaphne pendulifolia</i> Gamb. (KL 5250)	80
4.1.2	Isolation of Alkaloid from the Barks of <i>Alseodaphne pendulifolia</i> Gamb. (KL 5732)	81





LIST OF CHEMICAL COMPOUNDS STRUCTURE

No.	Chemical compounds structure name	Page
1	Citric acid	5
2	Tartaric acid	5
3	Malic acid	5
4	Cysteine	5
5	Tryptophan	5
6	Histidine	5
7	Serine	5
8	Alanine	5
9	Glutamic acid	6
10	Vanillin	6
11	Anthocyanins	6
12	Resveratrol	6
13	Morphine	6
14	Aspirin	6
15	Etoposide	6
16	Pyrethrins	6
17	Allethrins	6
18	Rotenone	7
19	Gibberellins	7
20	Cytokinins	7
21	Abscisic acid	7
22	Ethylene	7
23	Paclitaxel	20
24	Vitamin A	20
25	Vitamin D	21
26	Cantharidin	21
27	Doxorubicin	21
28	Tubocurarine	22
29	Cocaine	23
30	Codeine	23
31	Quinidine	23
32	Quinine	23
33	Vinblastine	23
34	L-ornithine	26
35	L-lysine	26
36	L-tyrosine	26
37	L-tryptophan	26
38	L-histidine	26
39	Usambarensine	26
40	Hordenine	26
41	Mescaline	26
42	Coniine	26
43	Capsaicin	27
44	Ephedrine	27
45	Solanidine	27
46	Caffeine	27



**LIST OF CHEMICAL COMPOUNDS STRUCTURE (conc.)**

47	Theobromine	27
48	Pinidine	27
49	(S)-Reticuline	29
50	Norglaucine	29
51	Palmatine	30
52	Buesgeniine	30
53	Allocryptopine	30
54	Thebaine	30
55	Emetine	30
56	(+)-Stephasubine	30
57	Dopamine	34
58	4-hydrophenylacetadehyde	34
59	(S)-Norcoclurine	34
60	Corytuberine	35
61	Isoboldine	35
62	Glaucine	35
63	Bulbocapnine	35
64	Papaverine	34
65	Berberine	34
66	Sanguinarine	34
67	Erythraline	34
68	Argemonine	34
69	(S)-cocclaurine	38
70	N-methylcocclaurine	38
71	Berbamunine	38
72	Dauricine	40
73	Isoliensinine	40
74	Pisopowiaridine	40
75	Imine cordobimine	40
76	Stebisimine	41
77	Sciadoferine	41
78	Nortiliacorinine A	41
79	Cissampareine	41
80	1,2-Dehydromicranthine	42
81	Pseudorepanduline	42
82	Insulanoline	42
83	Dihydroisoobtusilactone	46
84	Dihydroobtusilactone	46
85	3-Epilitsenolide D ₁	46
86	3-Epilitsenolide D ₂	46
87	Alseodafuranone	46
88	Sanjoinine	46
89	(-)-N-norarmepavine	46
90	(+)-Reticuline	46
91	(+)-Cocclaurine	47
92	(-)-Cocclaurine	47
93	Lirioferine	47





LIST OF CHEMICAL STRUCTURE (conc.)

94	Isocorydine	47
95	Norisocorydine	47
96	Obamegine	47
97	<i>N</i> -methyllaurotetanine	48
98	2-Norobamegine	48
99	Laurotetanine	48
100	(+)-2'-Noroxycanthine	48
101	(+)-Seeperine	48
102	(-)-Gyrolidine	49
103	(+)-Norstephasubine	49
104	(+)-2-Norobaberine	49
105	<i>O</i> -Methyllimacusine	49
106	Norboldine	50
107	(+)-3',4'-Dihydrostephasubine	50
108	<i>O</i> -Methylrepandine	50
109	Armepavine	50
110	Doryafranine	50
111	Xylopinine	51
112	1-(4-Methoxybenzyl)-6-7-methylenedioxy-1,2,3,4-tetrahydroisoquinoline	51
113	4-Hydroxy, 3-methoxy benzoic acid	51
114	Neolignan eusiderin-A	51
115	Norlirioferine	51
116	Lindcarpine	51
117	Norpredicentrine	51
118	α' -oxoperakensimine	52
119	Boldine	52
120	<i>N</i> -(2-hydroxypropyl)-pendulifoline A	52
121	<i>N</i> -(2-hydroxypropyl)-pendulifoline B	52
122	Oxohernagine	52
123	Hernagine	52
124	Corydine	52
125	Isocorytuberine	53
126	Costaricine	53
127	Sebiferine	53
128	<i>N</i> -(2-hydroxypropyl)-norboldine A	53
129	<i>N</i> -(2-hydroxypropyl)-norboldine B	53
130	<i>N</i> -cyanomethyl-norboldine	53
131	α' -oxoperakensimine A	54
132	α' -oxoperakensimine B	54
133	α' -oxoperakensimine C	54
134	<i>N</i> -methyl-2, 3, 6-trimethoxy morphinandien-7-one <i>N</i> -oxide	54
135	<i>N</i> -methyl-2, 3, 6-trimethoxymorphinandien- 7-one	54
136	Perakensol (7-hydroxy-2, 3, 6- trimethoxyphenanthrene)	55
137	Srilankine	55
138	1,1-Diethoxy-ethane	55
139	1-Methyl-1H-pyrrole	55





LIST OF CHEMICAL STRUCTURE (conc.)

140	1,1-Diethoxy-2-methyl-propane	55
141	2-Methoxy-phenol	55
142	4-One,2,3-dihydro-3,5-dihydroxy-6-methyl-4H-pyran	55
143	2-Methoxy-5-methylphenol	55
144	2-Methoxy-4-vinylphenol	55
145	D-allose	55
146	2,6-Dimethoxy-phenol	55
147	<i>N</i> -(2-morpholinoethyl)benzamide	55
148	1-Ethenyl-1-methyl-2,4-bis(1-methylethenyl)-cyclohexane	55
149	Sucrose	56
150	1,2,3,4-Tetrahydro-6,7-dimethoxy-2-methyl-isoquinoline	56
151	2-Methoxy-4-(1-propenyl)-phenol	56
152	3,4,5-Trimethoxy-phenol	56
153	3,5-Dimethyl-2-nitro-phenol	56
154	2-Nonadecanone	56
155	2,2-Dimethyl-cyclohexanol	56
156	Tridecanoic acid methyl ester	56
157	3',5'-Dimethoxyacetophenone	56
158	(<i>E</i>)-3-Eicosene	56
159	2-Benzoyl-4-methoxymethyl-6-methyl-furo[2,3- <i>b</i>]pyridin-3-amine	56
160	2-Oxo-hexadecanoic acid, methyl ester	56
161	5-Hydroxy-2,3,3-trimethyl-2-(3-methyl-buta-1,3-dienyl)cyclohexanone	57
162	Di(3,4,methylenedioxy)benzylamine acetate	57
163	1-Hexyl-2-nitrocyclohexane	57
164	2-Methyl-3-(1-methylethenyl)-,acetate,(1à,2à,3à)-cyclohexanol	57
165	(-)-1,2,3,4-Tetrahydro isoquinolin-6-ol-1-carboxylic acid, 7-methoxy-1-methyl-methyl ester	57
166	Norisocorydine <i>N</i> -oxide	82
167	Corydine <i>N</i> -oxide	85
168	Isocorydine <i>N</i> -oxide	85
169	<i>N</i> -methylhernagine	101
170	Hernovine	123





ABBREVIATIONS

^{13}C NMR	13-Carbon NMR
1D-NMR	One Dimension Nuclear Magnetic Resonance
^1H NMR	Proton NMR
2D-NMR	Two Dimension Nuclear Magnetic Resonance
CC	Column Chromatography
CD_3OD	Deuterated methanol
CDCl_3	Deuterated chloroform
CH_2Cl_2	Dichloromethane
CH_3	Methyl group
CHCl_3	Chloroform
cm^{-1}	Per centimetre
COSY	^1H - ^1H Correlation Spectroscopy
<i>D</i>	Doublet
<i>Dd</i>	Doublet of doublet
DEPT	Distortionless Enhancement by Polarization Transfer
DMSO	Dimethylsulphoxide
FT-NMR	Fourier Transform Nuclear Magnetic Resonance
G	Gram
HCl	Hydrogen chloride
HMBC	Heteronuclear Multiple Bond Correlation
HMQC	Heteronuclear Multiple Quantum Correlation
HREIMS	High Resolution Electron Spray Ionization Mass Spectroscopy



ABBREVIATIONS (conc.)

HRP2	Histidine-rich Protein 2
HSQC	Heteronuclear Single Quantum Correlation
Hz	Hertz
<i>J</i>	Coupling constant
kg	Kilogram
LC/MS	Liquid Chromatography – Mass Spectrometer
M	Molar
M	Meter
<i>m</i>	Multiplet
MeOH	Methanol
MHz	Mega Hertz
ml	Milliliter
mM	Milimolar
NH ₃	Ammonia
NMR	Nuclear Magnetic Resonance
OCH ₃	Methoxyl group
OH	Hydroxyl group
pH	Power of Hydrogen
Ppm	Parts per million
PTLC	Preparative Thin Layer Chromatography
s	Singlet

ABBREVIATIONS (conc.)

t	Triplet
TLC	Thin Layer Chromatography
α	Alpha
β	Beta
δ	Chemical shift
λ	Maximum wave length

CHAPTER 1

INTRODUCTION

1.1 General

Every single organism are made up by hundreds of thousands different chemical and each species producing its own characteristic mixture. Chemical composition in organism differ one to others, for example taste, scents, and colour of durian are totally different from apple because of its distinctive chemical composition. In spite of hundred or thousand of chemicals in the organism, each entities has their own ability of function either it was yet discovery or not by our modern scientists. But some different substances could share same properties (e.g., taste from citric acid **1** from citrus, tartaric acid **2** from grapes, and malic acid **3** from apples are perceived as sour by human tongue).



In daily lives, besides of the varieties of plants in the world, delicious taste of fruits are beneficial to human body in various ways due to their distinctive chemical composition. In the Holy Quran, a number of fruits; banana (al-Waqi'ah [56]:28-33), dates (ar-Ra'd [13]:4), olive (an-Nahl [16]:10-11), pomegranate (al-'An'am [6]:141), fig (at-Tin [95]:1) and grapes (al-Mu'minun [23]:19) have been proven by modern science have their own nutritious to prevent or cure various of disease (Al-Muammar & Khan, 2012; Biglari, AlKarkhi, & Easa, 2008; Bitler et al., 2007; Dueñas, Pérez-Alonso, Santos-Buelga, & Escribano-Bailón, 2008; Leifert & Abeywardena, 2008; Scarminio, Fruet, Witaicenis, Rall, & Di Stasi, 2012). In verses of al-'An'am ([6]:99), Allah S.W.T. commands human to observe and recount the variety and the beauty of His amazing creation with the obligation to protect sustainable of environment (Azlan & Azad, 2012; *Holy Quran: English Translation by Abdullah Yusuf Ali*, 2000).



وَهُوَ الَّذِي أَنْزَلَ مِنَ السَّمَاءِ مَاءً فَأَخْرَجْنَا بِهِ نَبَاتَ كُلِّ شَيْءٍ
فَأَخْرَجْنَا مِنْهُ خَضِرًا مَخْرُجًا مِنْهُ حَبًّا مُتَرَاكِبًا وَمِنَ النَّخْلِ
مِنْ طَلْعِهَا قِنْوَانٌ دَانِيَةٌ وَجَنَّاتٍ مِنْ أَعْنَابٍ وَالزَّيْتُونَ وَالرُّمَّانَ
مُشْتَبِهًا وَغَيْرَ مُتَشَبِهٍ انْظُرُوا إِلَى ثَمَرِهِ إِذَا أَثْمَرَ وَيَنْعِهِ إِنَّ فِي
ذَلِكَ لَآيَاتٍ لِقَوْمٍ يُؤْمِنُونَ ﴿٩٩﴾

It is He Who sendeth down rain from the skies: with it We produce vegetation of all kinds: from some We produce green (crops), out of which We produce grain, heaped up (at harvest); out of the date-palm and its sheaths (or spathes) (come) clusters of dates hanging low and



near: and (then there are) gardens of grapes, and olives, and pomegranates, each similar (in kind) yet different (in variety): when they begin to bear fruit, feast your eyes with the fruit and the ripeness thereof. Behold! in these things there are signs for people who believe (al-An'am [6]:99).

Each specific part of certain plant parts are produces numerous beneficial drugs products for mankind usage. Higher plants contribute about 25% of the total drugs in clinical use in the world. The specific plants organs that usually used likes herba or aerial parts (herba), leaf (folium), flower (flos), fruits (fructus), bark (cortex), root (radix), rhizome (rhizoma), and bulb (bulbus). The examples of variety plants parts that used as phytotherapeutic products are listed in Table 1.1.1(Skalicka-Wozniak, Widelski, & Glowniak, 2008).

Demand on natural products such as α -amino acid demands over two million tons per year to produce various products that important in industries such as cysteine 4, tryptophan 5, and histidine 6 as antioxidants; serine 7 and alanine 8 are used in cosmetics; and the widely used as flavor enhancer; glutamic acid 9 or commonly known as monosodium glutamate (MSG) (Ahluwalia, Kumar, & Kumar, 2006). Nowadays, natural products were not only restricted to food industries, but they are widely applied in other field of usage such as beverages (vanillin 10, anthocyanins 11, and resveratrol 12), pharmaceuticals (morphine 13, aspirin 14, and etoposide 15), pesticides (pyrethrins 16, allethrins 17, and rotenone 18) and plant growth regulators (gibberellins 19, cytokinins 20, abscisic acid 21 and ethylene 22)(Chinou, 2008).

Table 1.1.1

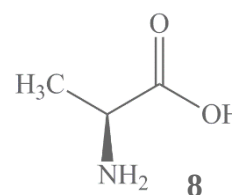
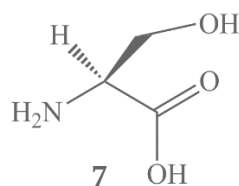
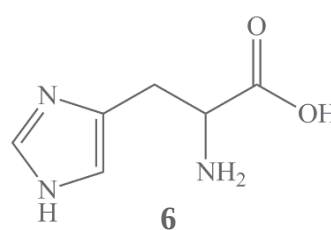
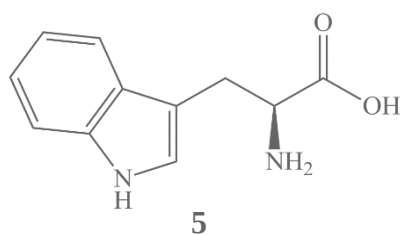
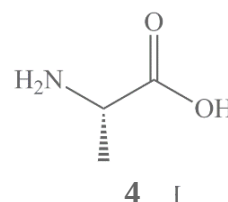
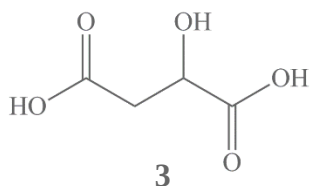
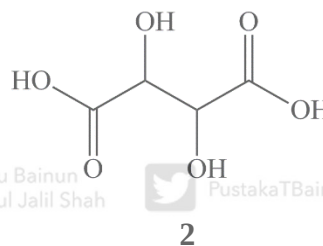
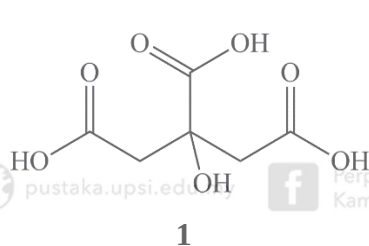
Specific Plants Parts in Certain Plants Species that Used in Phytotherapy.

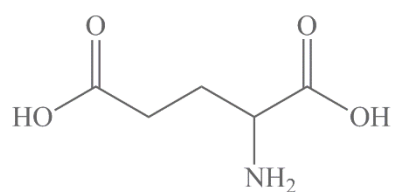
Plants parts	Plants
Fruits and seeds	Caraway (<i>Carum carvi</i>) Fennel (<i>Foeniculum vulgare</i>) Saw palmetto (<i>Serenoa repens</i>) Horse chestnut (<i>Aesculus hippocastanum</i>)
Leaf	Balm (<i>Melissa officinalis</i>) Deadly nightshade (<i>Atropa belladonna</i>) Ginkgo (<i>Ginkgo biloba</i>) Peppermint (<i>Mentha piperita</i>) Bearberry (<i>Arctostaphylos uva-ursi</i>)
Flowers	Chamomile (<i>Chamomilla recutita</i>) Calendula (<i>Calendula officinalis</i>) Arnica (<i>Arnica Montana</i>)
Stem	Ephedra (<i>Ephedra sinica</i>) Hawthorn (<i>Crataegus monogyna</i>) Passion flower (<i>Passiflora incarnata</i>) Wormwood (<i>Artemisia absinthium</i>)
Bark	Frangula (<i>Rhamnus frangula</i>) Willow (<i>Salix alba</i>)
Rhizome and root	Devil's claw (<i>Harpagophytum procubens</i>) T tormentill (<i>Potentilla erecta</i>) Rhubarb (<i>Rheum palmatum</i>) Kava-kava (<i>Piper methysticum</i>)

1.2 Problem Statements

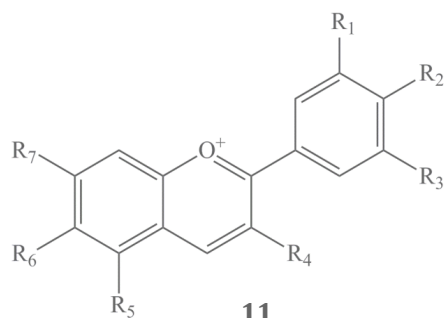
Malaysia is a tropical rain forest country which has been blessed with plethora of natural resources. Stand by its own, North Borneo indicated 3500 species of endemic plant which exhibiting high levels vascular plant species richness and endemism, while the other 2400 species thrives in Malaysian Peninsula (Ahmad et al., 2010; J.

Williams et al., 2001). Today, illegal logging activities for agriculture practice in our forest was endangered the flora and the might be extinct within time before our phytochemist and pharmacist could discover their chemical constituents and potential as our new drugs discovery. A fast action must be taken to overcome this problem. This study will provide a new literature about alkaloid constituents and their biological activities from *Alseodaphne pendulifolia* Gamb. that collected from two different locations. This is the first report that study of alkaloid constituents from bark of this species. From the literature review, not much papers are reported about chemical constituents from genus of *Alseodaphne* (Thakur et al., 2012).

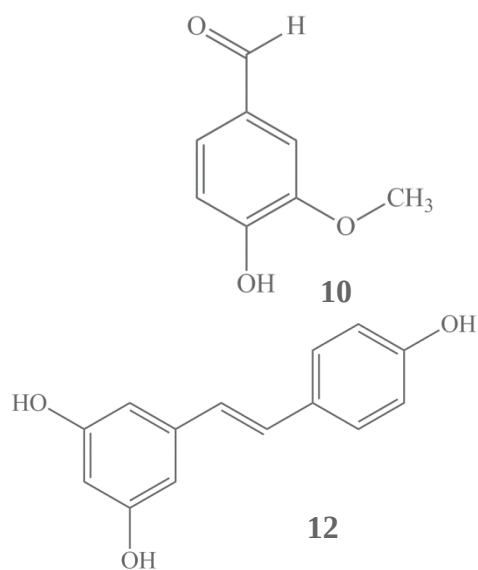




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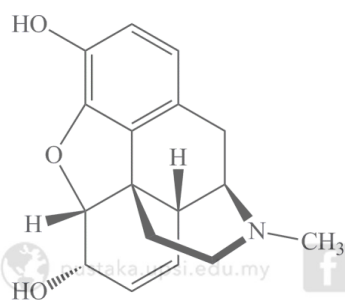


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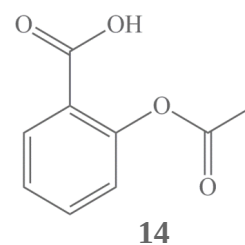


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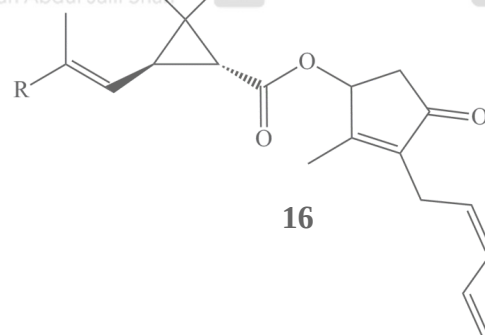
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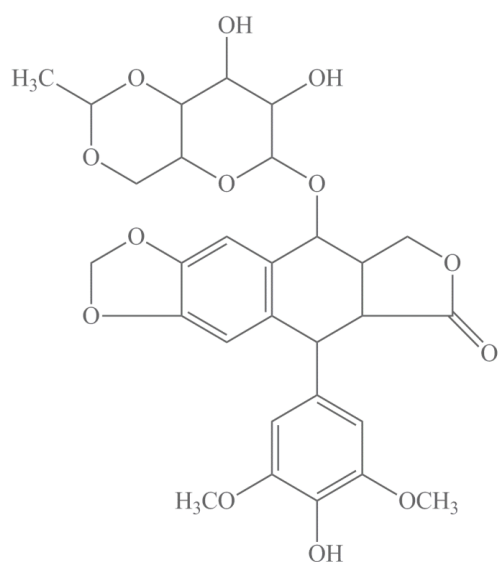
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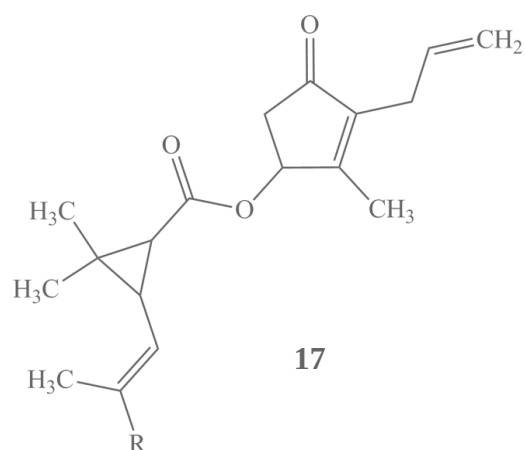
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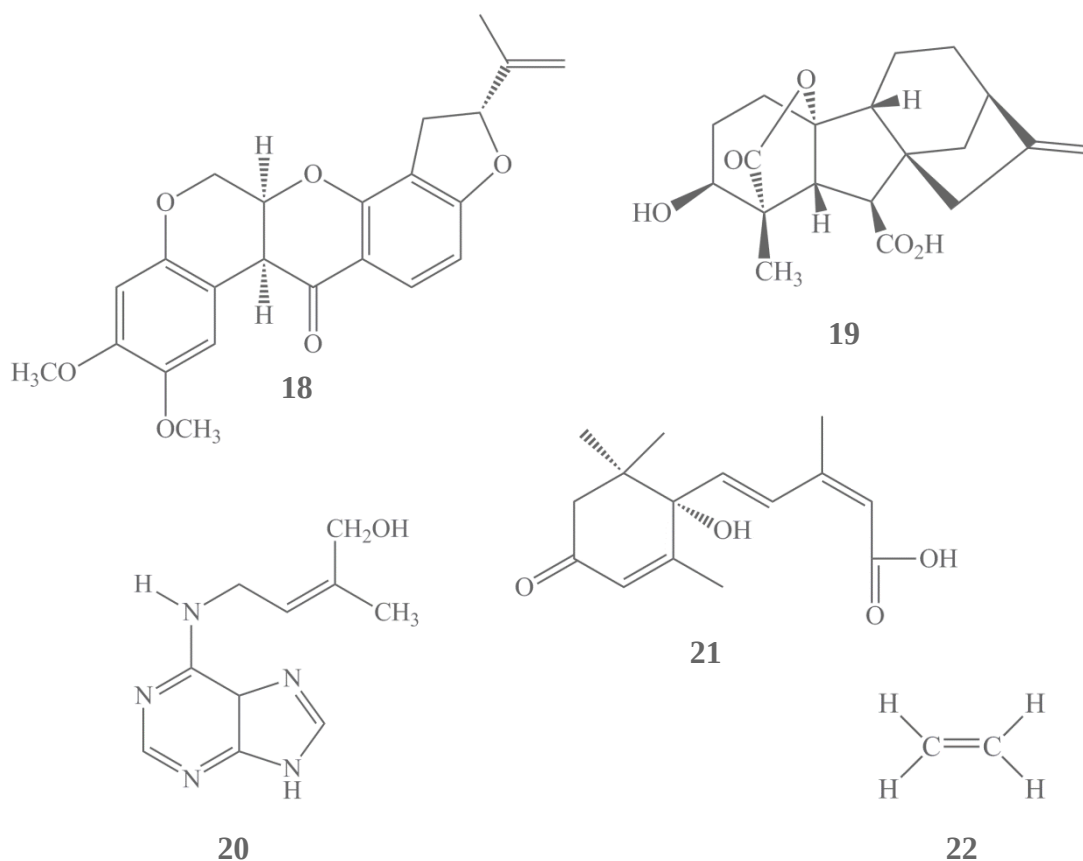


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1.3 Objectives of Study

The objectives of the study are as follows:

- To extract, isolate and purify alkaloid constituents from barks of *Alseodaphne pendulifolia* Gamb.
- To identify and elucidate the structure of the isolated alkaloids constituent using modern spectroscopy methods.
- To determine the antiplasmodial, cytotoxicity, antibacterial and antioxidant activities against the isolated alkaloids from the barks of *Alseodaphne pendulifolia* Gamb.



1.4 Family of Lauraceae

The Lauraceae are predominantly arboreous family, a large pantropical family mainly composed of large trees or shrubs that mostly found in warm, tropical, humid subtropical and mild temperate regions of northern and southern hemispheres (Joshi, Verma, & Mathela, 2010; Simie et al., 2004). Sixty eight genera and more than two thousand species have been reported under this family (Drewes, Horn, & Mavi, 1997; Kumar, Rawat, Rahuja, Srivastava, & Maurya, 2009; Little, Stockey, & Penner, 2009; Oh et al., 2012; “The plant list, a working list of all known plant species.,” 2010). The taxonomic hierarchy of Lauraceae family have been enlisted below:

Domain : Eukaryota



Kingdom : Plantae



Division : Magnoliophyta

Class : Magnoliopsida

Order : Laurales

Family : Lauraceae

Genera :

- | | |
|-----------------|--------------------|
| 1. Actinodaphne | 7. Beilschmiedia |
| 2. Aiouea | 8. Camphora |
| 3. Alseodaphne | 9. Caryodaphnopsis |
| 4. Aniba | 10. Cassytha |
| 5. Apollonias | 11. Chlorocardium |
| 6. Aspidostemon | 12. Cinnadenia |





13. Cinnamomum

14. Cryptocarya

15. Dehaasia

16. Dicypellium

17. Dodecadenia

18. Endiandra

19. Endlicheria

20. Eusideroxylon

21. Gamanthera

22. Hufelandia

23. Hypodaphnis

24. Iteadaphne

25. Kubitzkia

26. Laurus

27. Licaria

28. Lindera

29. Litsea

30. Machilus

31. Malapoenna

32. Mespilodaphne

33. Mezilaurus

34. Mocinnodaphne

35. Mutisiopersea

36. Nectandra

37. Neocinnamomum

38. Neolitsea

39. Nesodaphne

40. Notaphoebe

41. Nothaphoebe

42. Ocotea

43. Oreodaphne

44. Paraia

45. Parasassafras

46. Parthenoxylon

47. Persea

48. Phoebe

49. Phyllostemonodaphne

50. Pleurothyrium

51. Polyadenia

52. Potameia

53. Potoxylon

54. Povedadaphne

55. Ravensara

56. Rhodostemonodaphne

57. Sassafras

58. Schauera

59. Sextonia

60. Sinopora

61. Sinosassafras

62. Syndiclis



63. Systemonodaphne

66. Urbanodendron

64. Tetranthera

67. Williamodendron

65. Umbellularia

68. Yasunia

This family is largely characterized by trimerous flowers, bi- or tetrasporangiate anthers with apical valvate dehiscence, and a unilocular gynoecium containing a single anatropous, apically attached ovule. Fruits of the family are fleshy, typically black to almost black at maturity and usually borne on enlarged receptacles that can form cupulate to fully enclosing structures. Fruits of Lauraceae are typically considered drupes, but are better classified as a berry since the “endocarp” is only made up of a thin, single-celled sclerified endodermis. Oil or mucilage idioblasts occur throughout the plant tissues, including the wood (Little et al., 2009).

Name of Lauraceae itself is derived from the prominent member, the Grecian laurel, *Laurus nobilis*, is characterized by plants which have prominent oil cells in the leaves, wood and fruits. These oils are mostly aromatic hence provide a number of flavouring materials and spices (Schroeder, 1975). Recorded showed that the lauraceous plants have widespread use in both oriental medicine and occidental medicine (Shimonura, Sashida, Mimaki, Oohara, & Fukai, 1988). Some plants in this family have been employed in folk medicine for their interesting bioactivities (Rachmatiah, Mukhtar, Nafiah, Hanafi, Awang, et al., 2009; Rachmatiah, Mukhtar, Nafiah, Hanafi, Kosela, et al., 2009). *Cinnamomum camphora* (L.) Presl is a major source of camphor, which can be made into camphor oil and mothballs. In addition, camphor is taken orally to calm hysteria, nervousness, neuralgia and to treat serious diarrhoea. Camphor is also known to be effective in treating colds and chills (Lee et

al., 2006). The extracts from *C. Cassia* have been claimed to reduce inflammation (Lee & Shibamoto, 2002) and to decrease serum glucose, total cholesterol and platelet counts (Khan, Safdar, Ali-Khan, Khattak, & Anderson, 2003). Table 1.3.1 showed uses of species from family of Lauraceae.

Table 1.3.1

Uses of Species from Family of Lauraceae

Uses	Species of Lauraceae	References
Spices	<i>Cinnamomum cassia</i>	(Gottlieb, 1972; Kumar et al., 2009;
	<i>Cinnamomum subavenium</i>	Perera & Rajapakse, 1991)
	<i>Cinnamomum zeylanicum</i>	
	<i>Laurus nobilis</i>	
	<i>Sassafras albidum</i>	
Nutritious fruits	<i>Persea americana</i>	(Gottlieb, 1972; Kumar et al., 2009;
	<i>Persea gratissima</i>	Perera & Rajapakse, 1991)
Chinese folk medicines	<i>Cinnamomum cassia</i>	(Kumar et al., 2009)
	<i>Lindera aggregate</i>	
Timber	<i>Eusideroxylon zwageri</i>	(Gottlieb, 1972)
	<i>Mezilaurus itauba</i> ,	
	<i>Mezilaurus navalium</i>	
	<i>Ocotea bullata</i>	
	<i>Persea nanmu</i>	
Seed fat	<i>Laurus nobilis</i>	(Gottlieb, 1972)
	<i>Litsea sebifera</i>	
Perfume oils	<i>Aniba duckei</i>	(Gottlieb, 1972)
	<i>Aniba rosaeodora</i>	
	<i>Ocotea pretiosa</i>	

1.5 Genus of *Alseodaphne*

Genus of *Alseodaphne* consist more than fifty species (Table 1.4.1), distributed around Malaysia, China, Cambodia, Vietnam, Laos, Indonesia, Sri Lanka, Myanmar, Philippines, Thailand and India. This genus locally known as medang or tejur in Malaysia (International Seed Testing Association, 2013; Omar, Nafiah, Mukhtar, Awang, & Hadi, 2009), Wewarana in Sri Langka (Ariyadasa, 2002), Gemur in Indonesia (Joosten, Tapio-Bistrom, & Tol, 2012; Kalimantan Forests and Climate Partnership, 2009) and Nelthare in Tamil Arambamaram, Kanaippirandai (Alex, Charles, Leo, Joseph, & Mani, 2012; Charles, Stanly, Joseph, & Ramani, 2011).

From more than sixty species, only a few from this species were reported their usage. The trees of *Alseodaphne semecarfolia* was reported used as nest by Hornbill (Mudappa & Kannan, 1997; Velho, Datta, & Isvaran, 2009) and fruits was eaten as one of their food sources in Sg. Tekam Forest Reserve (Andrew D. Johns, 1987) but for *Alseodaphne perakensis*, aborigines of the Sakai tribe claimed that the fruit from this plant is poisonous (Lajis, Mahmud, Din, & Toia, 1989). The versatile and aesthetic qualities hardwood of *Alseodaphne hainanensis*, *A.oblanceolata* and *A. perakensis*, are highly demand in specific market of “luxury” applications including buildings, furniture, ships and ornaments due to their quality of hue, grain, texture, strength and durability (Huang, 2011; Lajis et al., 1989; Schroeder, 1982). In construction sector, the durability of *A. dura* and *A. longipes* is leveraged as seawater piles (Schroeder, 1982).

In Sumatra, Indonesia, *A. coriacea* is a livelihood sources for some Sumatran. It was harvested in deep forest for their valuable bark that rich with pyrethrins **16**, a compound that used in variety of insecticides especially in production of mosquito coil. Besides insecticides, the bark of this species also used in production of joss sticks (dupa cina), surfactant for recycling engine oil and some in cosmetics products (Janudianto, Mulyoutami, Joshi, Wardell, & Noordwijk, 2011; Joosten et al., 2012; Kristedi & Kieft, 2010; Lyons, 2003). In folk medicine, *A. semicarpifolia* was used as treatment of some illness likes cholera-like illness, rinderpest and dysentery diseases in cattle and also bark juice is applied externally for leach bite or chest pain (Alex et al., 2012; *Basic Medical Sciences; Intramural Research: An Annual Report*, 2012; Charles, Joseph, & Ramani, 2012; Charles et al., 2011; Harsha, Shripathi, & Hegde, 2005; Karuppusamy, 2007; Parinitha, Harish, Vivek, Mahesh, & Shivanna, 2004)

Alseodaphne are classified as evergreen trees. The leaves were alternate, always clustered near apex of branchlet, pinninerved, often turning black when dry. Inflorescence axillary, paniculate or racemose; bracts and bracteoles deciduous. Flowers are bisexual and trimerous. Perianth tube short; perianth lobes, subequal or outer is smaller, slightly dilated after anthesis but absent in fruit. Fertile stamens, in three whorls; filaments of first and second whorls glandless, those of third whorl each with two glands at base; anthers four-celled; cells of first and second whorls introrse, those of third whorl extrorse or upper two lateral and lower two extrorse. Staminodes of innermost whorl, very small, nearly sagittate. Ovary partly immersed into shallow perianth tube; style often as long as ovary; stigma small, inconspicuous, discoid. Fruit black or purplish black when mature, ovoid, oblong, or subglobose; fruit stalk red,



green, or yellow, sometimes nearly cylindric, fleshy, pulpy, always warty, truncate at apex (Shu, Hsi-wen, Jie, & Werff, 2008).

Table 1.4.1

List of Species in Genus of Alseodaphne (Lauraceae)

No.	Species	References
1	<i>A. albifrons</i>	I,II,III
2	<i>A. andersonii</i>	II,IV,V
3	<i>A. archboldiana</i>	VI
4	<i>A. bancana</i>	II,VII,VIII
5	<i>A. birmanica</i>	II
6	<i>A. borneensis</i>	II
7	<i>A. cavaleriei</i>	II
8	<i>A. ceratoxylon</i>	IX,X
9	<i>A. changchangensis</i>	IV
10	<i>A. coriacea</i>	I,XI,XII
11	<i>A. corneri</i>	II,XIII
12	<i>A. costalis</i>	VIII
13	<i>A. cuneata</i>	X
14	<i>A. dura</i>	II,XIV
15	<i>A. elmeri</i>	II
16	<i>A. elongata</i>	II
17	<i>A. falcate</i>	X
18	<i>A. foxiana</i>	II





19	<i>A. garciniicarpa</i>	II
20	<i>A. gigaphylla</i>	IV
21	<i>A. glauciflora</i>	II
22	<i>A. glaucina</i>	II
23	<i>A. gracilis</i>	II,V
24	<i>A. grandis</i>	VIII
25	<i>A. griffithii</i>	II
26	<i>A. habrotricha</i>	II
27	<i>A. hainanensis</i>	II,IV,V,XV,XVI,XVII
28	<i>A. havilandii</i>	X
29	<i>A. himalayana</i>	II
30	<i>A. hokouensis</i>	II,IV,V
31	<i>A. huanglianshanensis</i>	II,IV,V
32	<i>A. insignis</i>	I,II,IX,X,XVIII
33	<i>A. intermedia</i>	I,II,VII,XIX
34	<i>A. khasyana</i>	II
35	<i>A. kochummenii</i>	II
36	<i>A. languginosa</i>	XIV
37	<i>A. lanuginosa</i>	II
38	<i>A. longipes</i>	II,XIV,XVIII
39	<i>A. marlipoensis</i>	II,IV,V
40	<i>A. medogensis</i>	II
41	<i>A. micrantha</i>	II
42	<i>A. montana</i>	II





43	<i>A. nigrescens</i>	II,IV,VII,XX
44	<i>A. oblanceolata</i>	II,XIV,XIX
45	<i>A. obovata</i>	I
46	<i>A. owdenii</i>	II
47	<i>A. paludosa</i>	II,X
48	<i>A. peduncularis</i>	I,II
49	<i>A. pendulifolia</i>	I,II
50	<i>A. perakensis</i>	II,XIII,XIX,XXI,XXII,XXIII,XXIV
51	<i>A. petiolaris</i>	II,IV,V,XXV,XXVI,XXVII
52	<i>A. philippinensis</i>	II
53	<i>A. ramosii</i>	II
54	<i>A. rhododendropsis</i>	II
55	<i>A. ridleyi</i>	I,II
56	<i>A. rubriflora</i>	II
57	<i>A. rubrolignea</i>	II
58	<i>A. rugosa</i>	II,IV,V,XXVIII
59	<i>A. semecarpifolia</i>	II,IV,XXI,XXIX,XXX,XXXI,XXXII,XXXIII,XXIV,XXXV,XXXVI,XXXVII,XXXVIII,XXXIX,XL,XLI
60	<i>A. siamensis</i>	II
61	<i>A. sichourensis</i>	II,IV,V
62	<i>A. suboppositifolia</i>	II
78	<i>A. sulcata</i>	II
64	<i>A. tomentosa</i>	II
65	<i>A. tonkinensis</i>	II



66	<i>A. utilis</i>	II
67	<i>A. umbaliflora</i>	XVIII,XLII
68	<i>A. wrayi</i>	II
69	<i>A. yunnanensis</i>	II,V

References : I (International Seed Testing Association, 2013), II (Thakur et al., 2012), III (Zulkifly, Kim, Majid, & Merican, 2011), IV (Li et al., 2011), V (Shu et al., 2008), VI (Bick, 1996), VII (Chong, Tan, & Corlett, 2009), VIII (Wong et al., 2013), IX (Kartawinata et al., 2006b), X (Kartawinata et al., 2006a), XI (Joosten et al., 2012), XII (Kalimantan Forests and Climate Partnership, 2009), XIII (Sulaiman et al., 2011), XIV (Schroeder, 1982), XV (Huang, 2011), XVI (Ortega et al., 2010), XVII (Zhang & Ma, 2008), XVIII (Hidayat & Simpson, 1994), XIX (Ya'akub, Kudus, Nazre, Noor, & Hanum, 2013), XX (Tata, Noordwijk, & Werger, 2008), XXI (Lajis et al., 1989), XXII (Mahmud, 1989), XXIII (Nafiah et al., 2011), XXIV (Okechukwu, 2005), XXV (Lan, Zhu, & Cao, 2012), XXVI (Majumdar, Datta, & Shankar, 2012), XXVII (Rout, Sajem, & Nath, 2012), XXVIII (Koskela, Hong, & Rao, 2002), XXIX (Alex et al., 2012), XXX (Ariyadasa, 2002), XXXI (Chanderbali, Werff, & Renner, 2001), XXXII (Charles & Alex, 2011), XXXIII (Charles et al., 2012), XXXIV (Charles et al., 2011), XXXV (Harsha et al., 2005), XXXVI (Jayakumar, Arockiasamy, & Britto, 2000), XXXVII (Karuppusamy, 2007), XXXVIII (Mahadkar, Valvi, & Jadhav, 2013), XXXIX (Mudappa & Kannan, 1997), XL (Parinitha et al., 2004), XLI (Parthasarathy, 1999), and XLII (Adinugroho, Suryadiputra, Saharjo, & Siboro, 2011)

1.6 Species of *Alseodaphne pendulifolia* Gamb.

Alseodaphne pendulifolia Gamb. (Figure 1.5.1 – 1.5.3) is a medium to large sized tree with approximately 120 cm trunk circumference, height up to 18 m tall, that are rarely found in lowland forests around Perak, Terengganu, Pahang, Selangor, and Borneo Island. This plant has pale yellow flowers in auxiliary panicles, with perianth glabrous outside and outer whorl smaller than inner. The shape of fruits are obvoid with 1.5 x 1 cm in sized with about 15 cm long perianth tube which slightly enlarged (Nafiah, 2009; Ng, 1989; Thakur et al., 2012).

The colour of the bark is brownish, smooth to lenticels parts and the inner bark are yellow same as sapwood. The twigs are thick with prominent leaf scars. The leaves are clustered at the ends of the twigs, dropping down with stalk about 2 - 4 cm along and blade leathery, slightly glaucous below, obovate or oblanceolate shape. The apex is blunt or pointed and the base is narrowed. Midrib rose above and the secondary nerves are about 15-22 pairs, curving and joining near margin, which is visible on above side (Nafiah, 2009; Ng, 1989; Thakur et al., 2012).



Figure 1.5.1: Fruits of *Alseodaphne pendulifolia* Gamb.



Figure 1.5.2: Barks of *Alseodaphne pendulifolia* Gamb.



Figure 1.5.3: Leaves of *Alseodaphne pendulifolia* Gamb.