





EARLY DEVELOPMENT AND LARVAL REARING OF CLIMBING PERCH, Anabas testudineus Bloch



Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

October 2012











Abstract of thesis presented to the senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Masters of Science

EARLY DEVELOPMENT AND LARVAL REARING OF CLIMBING PERCH, Anabas testudineus Bloch

By

ZALINA BINTI ISMAIL

October 2012

Chair: Associate Professor Che Roos Bin Saad, PhD

Faculty: Agriculture

Anabas testudineus or locally known as 'puyu' is a freshwater fish species grown in Southeast Asian countries. This study was carried out from March – December 2010 at Aquaculture Experimental Station of Universiti Putra Malaysia. This fish is also known as a species that has a low survival rate during its early life stage and fry. Its Seed production and stock assessment are still poorly understood due the high mortality at first stage of development. In the rearing aspect, high food convertion ratio has been recorded when this fish is reared in hapas and earth ponds using homemade food.

The objectives of this study were to induce breed climbing perch (*Anabas testudineus*) using a commercial hormone preparation (Luteinizing Hormone Releasing Hormone analogue (LHRHa), to observe and record the morphological embryonic development of the *A. testudineus* and to determine optimal stocking density. The first experiment was conducted to







determine its the effectiveness of LHRHa as an agent to induce maturation and ovulation of A. testudineus with the intensity level of 2, 20, 200 µg/kg of body weight and saline as a control. The brooder were injected one time and left to spawn in the aquarium tanks in the sex ratio between male and female as 1:1. The parameters observed include fertilization rate, hatching rate, latency period, eggs production and oocytes diameter. For induced breeding, it was found that all intensity of LHRHa hormone level could enhance the fish to breed with the exception of the control group. It was observed that the fertilized eggs of A. testudineus were almost spherical in shape, clear pearl likes in appearance and free floating on water surface. Egg production was significantly higher in fish treated with 200 µg/kg as compared to fish treated with 2 and 20 µg/kg of body weight of LHRHa hormone while the highest hatching percentage (65.33%) was recorded in fish treated with 2 µg/kg of LHRHa hormone. There was no significant (P>0.05) effect between hormone level on fertilization rate and eggs diameter. The diameters of fertilized eggs ranged from 800 µm-850 µm.

For the second experiment, fertilized eggs were obtain through induced spawning and the development of embryos was monitored by sampling embryos at every 30 minutes to 1 h intervals until hatched. The first cleavage occurred at 1:30 h, epiboly began at 5 h, while the embryonic body was formed at 12 h and hatching occurred at 20 h after spawning at water temperature of 26°C.

Finally the third experiment was conducted to examine the effect of initial larval density of A. testudineus on growth and survival at three different





stocking densities of 35, 55 and 75 larvae/L. Newly hatched larvae of A. testudineus were produced by induced spawning using LHRHa hormone. Results showed that the survival and growth of A. testudineus larvae and fry during 28-day nursing period were stocking density dependent. The highest survival rate 75% was recorded in 35 larvae/L followed by 55 larvae/L (53%) and lastly 75 larvae/L (43%). Water quality parameters like temperature, pH, DO and ammonia ranged from 28.3±0.1°C to 28.3±0.3°C, 8.7±0.1 to 8.8±0.3, 5.7±0.6 to 5.8±0.4 ppm and 0.12±0.22 to 0.18±0.3 ppm respectively, were stable and not influenced by the stocking densities tested.

In conclusion, the use of LHRHa was proven to effectively induce maturation and ovulation in A. testudineus and the doses affected the eggs production and hatching rate.













Abstrak tesis yang dikemukakan kepada senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk jiazah Master Sains

PERKEMBANGAN AWAL DAN ASUHAN BENIH PUYU, Anabas testudineus Bloch

Oleh

ZALINA BINTI ISMAIL

Oktober 2012

Pengerusi: Profesor Madya Che Roos Bin Saad, PhD

Fakulti: Pertanian

Anabas testudineus atau nama tempatannya 'puyu' adalah spesies ikan air tawar yang membesar di Negara-negara Asia Tenggara. Kajian ini dijalankan pada Mac-Disember 2010 di Stesen Penyelidikan Akuakultur, Universiti Putra Malaysia. Ikan ini juga dikenali sebagai spesies yang mempunyai kadar hidup yang rendah di peringkat awal kehidupan menyebabkan pembiakan anak benih tidak dapat memenuhi permintaan pasaran. Kajian terhadap hasil pengeluaran anak benih dan bekalan stok masih lagi kurang dikaji kerana berhadapan dengan kadar kematian yang tinggi di awal peringkat pertumbuhan benih. Dalam aspek ternakan, nisbah pertukaran makanan yang tinggi telah dicatat apabila ikan diternak dalam hapa dan kolam tanah dengan menggunakan makanan buatan sendiri.

Objektif kajian ini adalah untuk merangsang pembiakan climbing perch (Anabas testudineus) menggunakan penyediaan hormon komersial LHRHa,





untuk memerhati dan merekodkan morfologi perkembangan embrio A, testudineus dan mengenalpasti kepadatan stok yang sesuai untuk meningkatkan kadar hidup.

Eksperimen pertama telah dijalankan untuk menentukan keberkesanan LHRHa sebagai agen yang menpercepatkan kematangan dan ovulasi A. testudineus dengan kepekatan hormon 2, 20 dan 200 µg/kg berat badan dan larutan salin sebagai kawalan. Induk telah disuntik dengan satu suntikan dan dibiarkan untuk bertelur dalam akuarium dengan kadar nisbah jantina jantan dan betina ialah 1:1. Parameter yang diteliti dalam eksperimen ini adalah kadar persenyawaan, kadar penetasan, tempoh matang ikan, jumlah telur setiap ikan dan diameter oosit. Bagi pembiakan aruhan, didapati semua kepekatan hormon LHRHa boleh merangsang pembiakan ikan kecuali kumpulan kawalan. Juga didapati, telur A. testudineus yang disenyawakan adalah dalam bentuk yang hampir sfera, mempunyai warna keputihan mutiara dan terapung dipermukaan air. Jumlah telur setiap ikan ketara lebih tinggi dalam ikan yang disuntik dengan kepekatan hormon 200 µg/kg berbanding dengan ikan yang disuntik dengan hormon LHRHa sebanyak 2 µg/kg dan 20 µg/kg daripada berat badan. Sementara itu, kadar penetasan yang tinggi direkod pada ikan yang disuntik dengan kepekatan hormon LHRHa 2 μ g/kg. Kesan antara dos hormon adalah tidak signifikan (P>0.05) untuk kadar persenyawaan dan diameter telur. Diameter telur yang disenyawakan berukuran antara 800-850 µm.





Bagi eksperimen kedua, persenyawaan telah dijalankan melalui pembiakan aruhan dan perkembangan embrio dipantau melalui persampelan embrio pada selang masa 30 minit ke 1 jam sehingga menetas. Pembahagian pertama sel telur berlaku pada 1:30 jam, epiboli bermula pada 5 jam, badan embrionik terbentuk pada 12 h dan penetasan berlaku pada 20 jam selepas bertelur pada suhu air 26°C.

Eksperimen ketiga ialah mengkaji kesan kepadatan larva semasa asuhan terhadap pertumbuhan dan kadar hidup A. testudineus pada tiga kepadatan stok berbeza iaitu 35, 55 dan 75 larva/L. Larva A. testudineus yang baru menetas dihasilkan daripada pembiakkan aruhan menggunakan hormon LHRHa 2 uq/kg. Hasil kajian menunjukkan bahawa kemandirian dan pertumbuhan A. testudineus dalam 28 hari tempoh asuhan adalah bergantung kepada kepadatan stok. Kadar hidup paling tinggi, 75% dicatatkan pada 35 larva/L, diikuti oleh 55 larva/L (53%) dan akhir sekali 75 larva/L (43%). Parameter kualiti air seperti suhu, pH, DO dan ammonia 28.3±0.1°C hingga 28.3±0.3°C, 8.7±0.1 hingga 8.8±0.3, 5.7±0.6 adalah hingga 5.8±0.4 ppm dan 0.12±0.22 hingga 0.18±0.3 ppm tiap satunya adalah stabil dan tidak dipengaruhi oleh kepadatan stok yang diuji.

Kesimpulannya, hormon LHRHa telah menunjukkan kesan afektif untuk merangsang kematangan dan peneluran pada A. testudineus dan kadar dos juga menunjukkan kesan pada jumlah telur setiap ikan dan kadar penetasan.







ACKNOWLEDGEMENTS

Foremost, I would like to express my sincere gratitude to my supervisor Assoc. Prof. Dr. Che Roos Saad for the continuous support of my master study and research, for his patience, motivation, enthusiasm, and immense knowledge. His guidance helped me in all the time of research and writing of this thesis. Thanks for generously providing guidance on the technical aspect of this thesis, for continuously encouraging me and pushing me to my limits to complete my thesis this semester, and for all the patience and support you gave me since day 1 of my thesis completion. I could not have imagined having a better supervisor and mentor for my master study.

I would like to express my deep and sincere gratitude to my co-supervisor, Assoc. Prof. Dr. Sharr Azni Harmin and Dr. Annie Christianus. Their wide knowledge and their logical way of thinking have been of great value for me. Their understanding, encouraging and personal guidance have provided a good basis for the present thesis. I owe my most sincere gratitude to Mr. Abdullah Abdul Rahim, for valuable advice and friendly helps. His extensive discussions around my work and interesting explorations in operations have been very helpful for this study.

I would like to extend my thanks to the staff at Aquaculture Research Station in Puchong, Selangor for their kindness in lending their hand during the experiments. Million thanks to Mr. Azmi Yaacob, Ms. Norazlina Nordin, Mr. Mohammad Syahrizan Shaharudin and Mr. Roszainal Yusop for their help.



Sincerely thanks to the Assistant Lab Officer, Mrs. Nur Shafika Maulad Abd Jalil, Mrs. Zaiton Basar and Mr. Jasni Mohd Yusoff for helping me out during the experiments.

I owe my loving thanks to my husband Shaikh Mohd Azhari Shaikh Yusof, without his encouragement and understanding it would have been impossible for me to finish this work. His support, encouragement, quiet patience and unwavering love were undeniably the bedrock upon which the past four years of my life have been built. His tolerance of my occasional vulgar moods is a testament in itself of his unyielding devotion and love. My special gratitude is due to my parent, my sisters and my mother in law for their loving support.

In my daily work I have been blessed with a friendly and cheerful group of fellow students. Sincere thanks to all my friends especially Noor Fazielawanie Mohd Rashid, Nurhidayu Al-saari, Nurul Ashikin Muhammad, Nik Md Azuadi Nik Daud, Norhidayah Mohd Taufek and Mohamad Faizul Mat Isa others for their kindness and moral support during my study. Thanks for the friendship and memories.

Last but not least I would like to express my gratitude to Ministry of Higher Education (MOHE) and Sultan Idris Education University (UPSI) for scholarship and financial support during my studies.



PustakaTBa

ptbupsi

I certify that a Thesis Examination Committee has met on 4 October 2012 to conduct the final examination of Zalina binti Ismail on her thesis entitled "Early Development and Larval Rearing of Climbing Perch, *Anabas testudineus* Bloch" in accordance with the Universities and University College Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

Mihdzar bin Abdul Kadir, PhD

Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Chairman)

Mohd Salleh bin Kamarudin, PhD

Professor Faculty of Agriculture Universiti Putra Malaysia (Internal Examiner)

Muta Harah binti Zakaria, PhD Associate Professor psiledu my Kampus Sultan Abdul Jalii Shah

Associate Professor estedumy Faculty of Agriculture Universiti Putra Malaysia (Internal Examiner)

Ahmed Jalal Khan Chowdhury, PhD

Professor Kulliyyah of Science International Islamic University Malaysia (External Examiner)

SEOW HENG FONG, PhD Professor and Deputy Dean School of Graduate Studies Universiti Putra Malaysia

Date: 21 March 2013

05-4









This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Che Roos Bin Saad, PhD Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Chairman)

Annie Christianus, PhD Lecturer Faculty of Agriculture Universiti Putra Malaysia (Member)

Sharr Azni Bin Harmin, PhD Professor Faculty of Science & Biotechnology Universiti Selangor (UNISEL) (Member)

BUJANG BIN KIM HUAT, PhD Professor and Dean School of Graduate Studies Universiti Putra Malaysia

1 1 APR 2013 Date:



DECLARATION

I declare that the thesis is my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously, and is not concurrently, submitted for any other degree at Universiti Putra Malaysia or at any other institution.

legalina

ZALINA BINTI ISMAIL

Date: 4 October 2012

















LIST OF TABLES

Table

Page

| 3.2.4 | Treatment and dosage for Anabas testudineus | 28 |
|---------|---|----|
| 3.3.1 | Oocytes diameter of female Anabas testudineus at different stage of ovulation in treatment (2, 20, 200 µg/kg) | 31 |
| 3.3.2 | Result of captive breeding experiments of <i>Anabas</i> testudineus treated with LHRH hormone | 33 |
| 4.3. | Embryonic characteristic and duration of development in Anabas testudineus | 42 |
| 5.3.2.1 | Survival rate and Specific Growth Rate of Anabas testudineus larvae nursed in aquaria filled with 2 liters of water for 28 days at three stocking densities | 66 |
| 5.3.2.2 | Weekly mean total length (mm) of <i>Anabas testudineus</i> nursed in aquaria filled with 2 liters of water for 28 days at three stocking densities | 67 |
| 5.3.2.3 | Weekly mean body weight (mg) of <i>Anabas testudineus</i> nursed in aquaria filled with 2 liters of water for 28 days at three stocking densities | 67 |
| 5.3.3 | Water quality parameters in nursing aquaria throughout the study period | 68 |





•



LIST OF FIGURES

Figure Page 2.1 Anabas testudineus Production in Malaysia 6 2.2.1 Map of Asia. Circle Showing the Distribution of Anabas 9 testudineus 2.2.4 Mature Male and Female of Anabas testudineus 12 2.6 Newly Hatched Larvae of Anabas testudineus 21 3.2.1 Genital Pore for Male and Female of Anabas 26 testudineus 3.2.3 Germinal Vesicle Stages of Anabas testudineus 27 Oocytes The Zygote Stage of Embryonic Development in 45 4.3.1 Anabas testudineus about 10 min after fertilization (scale bar = 0.2 mm).Vm–Vitelline membrane; Y- Yolk; Ps Perivitelline space. Figure show the dorsal views only 4.3.2 Cleavage Stage of Embryonic Development in Anabas 47 testudineus (scale bar = 0.2 mm). Panel (1a)-2 cell; (2a)-4 cell; (3a)-8 cell; (4a)-16 cell; (5a)-32 cell; (6a)-64 cell; (7a)-128 cell. Figure show the dorsal views only 48 4.3.3 Blastula Stage of Embryonic Development in Anabas testudineus (scale bar = 0.2 mm). Figure show the dorsal views only 4.3.4 Gastrula Stage of Embryonic Development in Anabas 50 testudineus (scale bar = 0.2 mm). (1)- Germ ring stage; (2)- 10% epiboly stage; (3)- 25% epiboly stage; (4)-50% epiboly stage; (5)-75% epiboly stage; gr-germ ring; eb- epiboly; p-polster; tb- tail bud; np-neural plate. Figure show the dorsal views only 4.3.5 Segmentation Stage (scale bar = 0.2 mm). (1)-2 52 somite; (2)-5 somite; (3)-6 somite; (4)-14 somite; (5)-15 somite (6)-20 somite; p-polster; np-neural plate; kvkupffer's vesicle; ov-optic vesicle; hb-hindbrain (rhombencephalon); fb-forebrain (prosencephalon); mb midbrain (mesencephalon); tb-tail bud; s-somite; cdcuvierian ducts. a- shows the dorsal view; b-shows the

lateral view





Hatching Stage (scale bar = 0.2 mm). Panel (1a)-4.3.6 54 Newly hatch larvae; (2a)-1st day larvae; (3a)-2 day old larvae; (4a)-3 day old larvae; (5a)-4 day old larvae; v.sbin-vt-subintestinal vitelline vein; v.crd.a-anterior cardinal vein; t-tail; pf-pelvic fin; uj-upper jaw; lj-lower jaw. a- show the lateral views only











LIST OF ABBREVIATIONS

| | BW | - | Body weight |
|---|------------|---------|--|
| | DO | - | Dissolved oxygen |
| | DOF | - | Department of Fisheries |
| | FAO | - | Food and Agriculture Organization |
| | FOM | - | Final oocytes maturation |
| | FSH | - | Follicular Stimulating Hormone |
| | GSI | - | Gonadosomatic Index |
| | GtH | - | Gonadotropin Hormone |
| | GV | - | Germinal Vesicle |
| | IM | - | Intramuscular injection |
| 3 | LH 4506832 | 😯 pūsta | ka Luteinizing Hormone Tuanku Bainun Kampus Sultan Abdul Jalii Shah |
| | LHRHa | - | Luteinizing Hormone Releasing |
| | | | Hormone analog |







TABLE OF CONTENTS

| APPROVAL DECLARAT LIST OF TA LIST OF FI | EDGEMENTS - FION ABLES | Page ii v viii x xii xiii xiv xvi |
|--|--|---|
| CHAPTER | | |
| 1 | INTRODUCTION | 1 |
| 2 | LITERATURE REVIEW 2.1 Aquaculture Industry 2.2 Biology of Climbing | 5 5 |
| | perch Anabas testudineus 2.2.1 Distribution 2.2.2 Common Names and Taxonomy 2.2.3 Morphological Characteristic 2.2.4 Sexual Dimorphism | 8 9 10 10 11 |
| | 2.3 2.4 Reproduction in Fish ank Bainin Gonad Development Abdul Jalil Shah 2.4.1 Gonad 2.4.2 Ovary | 13 15 15 15 |
| | 2.5 Hormone 2.5.1 Application of LHRHa in Induced breeding | 17 17 |
| | 2.6 Induced Breeding 2.7 The Larval 2.8 Effect of Stocking Density on Larval rearing | 19 20 21 |
| 3 | EFFECT OF LUTEINIZING HORMON RELEASING HORMONE (LHRHa) ON REPRODUCTIVE PERFORMANCE IN CLIMBING PERCH Anabas testudineus (BLOCH) 3.1 Introduction | 24 |
| | 3.2 Materials and Methods 3.2.1 Broodfish Selection 3.2.2 Hormone Preparation 3.2.3 Ovulatory Response | 25 25 26 27 |

- 3.2.4 Experimental Design
- 3.2.5Statistical Analysis30Results30
- 3.3 Results3.4 Discussion
- 3.5 Conclusion

05-4506

pustaka.u

Perpustakaan Tuanku Bainun Kampus Sultan Abdul Jalil Shah



28

34

37



| 4 | EMBRYONIC PERCH Anabas | DEVELOPMENT OF CLIMBING | | | |
|--|---|---|--|--|--|
| | 4.1 Introduct | | 38 39 39 39 40 | | |
| | 4.2.5 | Larval development | 41 | | |
| | 4.3 Results 4.3.1 4.3.2 4.3.3 4.3.4 4.3.5 4.3.6 | Zygote Period Cleavage Blastula Gastrula Segmentation Hatching | 41 45 46 48 49 51 53 | | |
| | 4.4 Discussion4.5 Conclusion | | 56 59 | | |
| 5 EFFECT OF STOCKING DENSITY ON GROWTH AND SURVIVAL IN CLIMBING PERCH Anabas testudineus | | | | | |
| | 5.1 Introduct | on Perpustakaan Tuanku Bainun and Methods ^{bodul Jalil Shah} Broodfish Selection Hormonal Injection Induced Spawning Larval Rearing | 60 61 61 62 62 63 | | |
| | 5.2.5 5.3 Results 5.3.1 5.3.2 5.3.3 | Statistical Analysis Breeding Performance Survival Rate and Growth Water Quality in Nursing aquaria | 64 65 65 65 68 | | |
| | 5.4 Discussion 5.5 Conclusion | | 69 71 | | |
| 6 | CONCLUSION | AND RECOMMENDATION | 72 | | |
| REFERENCES APPENDICES Appendix A: | | Feeding Schedule During Rearing | 74 87 87 | | |
| | Appendix B: | period Embryonic Development of <i>Anabas</i> | | | |
| Testudineus BIODATA OF STUDENT LIST OF PUBLICATION | | | | | |



CHAPTER 1 INTRODUCTION

Aquaculture is the farming of freshwater and marine organisms such as finfish, mollusks, crustaceans and aquatic plants (Parker, 2012). It involves cultivating aquatic populations under controlled or semi controlled conditions, and can be contrasted with commercial fishing, which harvests wild fish. For centuries, fish have been caught from sea, river and lake. Marine fish have been caught to the maximum level by the fisherman and this contributes to over exploitation. The demand of fish for food and ornamental aguaria is steadily increasing. Natural fish populations have declined during the last several decades because of environmental degradation and over-fishing (FAO, 2004). Due to this, aquaculture must be implemented seriously to supply and replace the caught fish protein demanded by the world population.

Induced spawning is very important in order to expand production of fish seed to meet the increased demand by fish farmers. Many fish spawn in environments that are nearly impossible to be simulated in a hatchery. Induced spawning by hormonal treatment is one of the ways to produce larvae or fry for stocking. Hormone induced spawning is the only reliable method to induce reproduction in most fishes. Hormone induced spawning of fish has been used for almost 60 years (Rotmann et al., 1991). Surprisingly, the same procedures, with only minor modifications, have been used to spawn an entire range of fishes from the ancient sturgeon and paddlefish to





carp, catfish, salmon, sea bass, redfish, snook, and mullet (Rotmann et al., 1991). In addition, induced spawning also can be used to produce hybrids that are different from the parent species, synchronize reproduction of large numbers of fish for simultaneous spawning, thereby simplifying production and marketing of the fish and produce fry outside the normal spawning season for maximum hatchery production (Rottmann et al., 1991). Bv applying this technique, farmers can provide fish when the price and market demand is greatest, and maximize the survival of fry under controlled hatchery conditions.

Studying the morphological and ecological features of eggs and larvae is important as it is a key issue for improving seed productivity as well as stock assessment (Morioka et al., 2009). Information on early embryonic and larval development is of critical importance in understanding the basic biology of a particular species and their dietary needs and environmental preferences (Borcato et al., 2004; Koumoundouros et al., 2001). Furthermore, studies on embryonic and early larval development are imperative and consequential to the successful rearing of larvae for large scale seed production and aquaculture (Rahman et al., 2004; Khan and Mollah, 1998). Fish embryonic development consists of seven stages leading to hatching. These stages are the zygote period, cleavage period, blastula period, gastrula period, segmentation period, pharyngula period, and finally hatching. There is a scarce literature of climbing perch (Anabas testudineus) on early embryonic development stage especially in Malaysia. Studies by Jalilah et al. (2011), Hughes et al. (1986), Singh and Mishra (1980) and Moitra et al. (1986) are

🕑 05-4506832 🛞 pustaka.upsi.edu.my 🚹 Perpustakaan Tuanku Bainun 💟 PustakaTBainun 🚺 ptbupsi





not that detailed and recent study on growth and morphology development on larvae have been reported by Morioka et al. (2009).

Various hormone applications have been performed on A. testudineus to stimulate maturation and ovulation such as ovaprim (Bhattacharyya and Homechaudhuri, 2009), LHRHa (Superfact) combination with a dopamine inhibitor (Motillium) (Morioka et al., 2009), Wova-FH (Sarkar et al., 2005) and heteroplastic pituitary gland extract (Moitra et al., 1986). All of these attempts have been successfully. However, a commonly used hormone (Superfact + dopamine inhibator), ovaprim and Wova-FH for larval production have some limitations due to the factor of high cost and their unsuitability to be injected in brood stock which can causing death. It is important to identify a hormone that is easy to be administered by farmers, more effective in little dose and can produce high production in order to reliably supply of larvae.

Luteinizing hormone releasing hormone (LHRHa) is the name of a mammalian hormone that has been employed successfully to induce the reproductive hormonal. In recent years, synthetic analogues of LHRHa have been developed that are far more effective. As they are purer and are not rapidly metabolized by fish, hormonal analog remains active for a longer period. The use of LHRHa for spawning induction has several advantages over the traditional hypophysation technique. The LHRHa is a small peptide molecule. Thus it can be synthesized into its native form and also into altered forms (analogues) with slow rates of degradation. A lower dose is required when using analogue forms (Zohar et al., 1989). LHRHa has been

🕑 05-4506832 🜍 pustaka.upsi.edu.my 🚹 Perpustakaan Tuanku Bainun 💟 PustakaTBainun 🚺 ptbupsi





successfully used for maturation and spawning of various fish including seabass and rabbit fish (Harvey et al., 1985) and Atlantic salmon (Crim et al., 1984).

Although efforts have been directed towards the morphological and ecological features of larval and juveniles stages, as key issues for improving seed productivity as well as stock assessment, these have not had the expected success (Chaco'n and Rosas, 1995), due to factors such as the high mortality during the first stages of development caused by the lack of knowledge involved in the development process. Therefore, morphological development can provide behavioural information as a part of ecology for larval and juvenile A. testudineus using laboratory reared specimen's data during the first weeks with the aim to develop techniques for the intensive culture of the species and to promote optimal management and conservation. Thus hypothesis of this study was use of LHRH hormone will increase the percentage of successful spawning in A. testudineus. Thus, the objectives for this work were:

- 1) To induce breed of Anabas testudineus using commercial hormone preparations (LHRHa).
- 2) To observe and record the embryonic development of Anabas testudineus.
- 3) To determine the effect of different stocking density on the growth and survival of Anabas testudineus larvae.

